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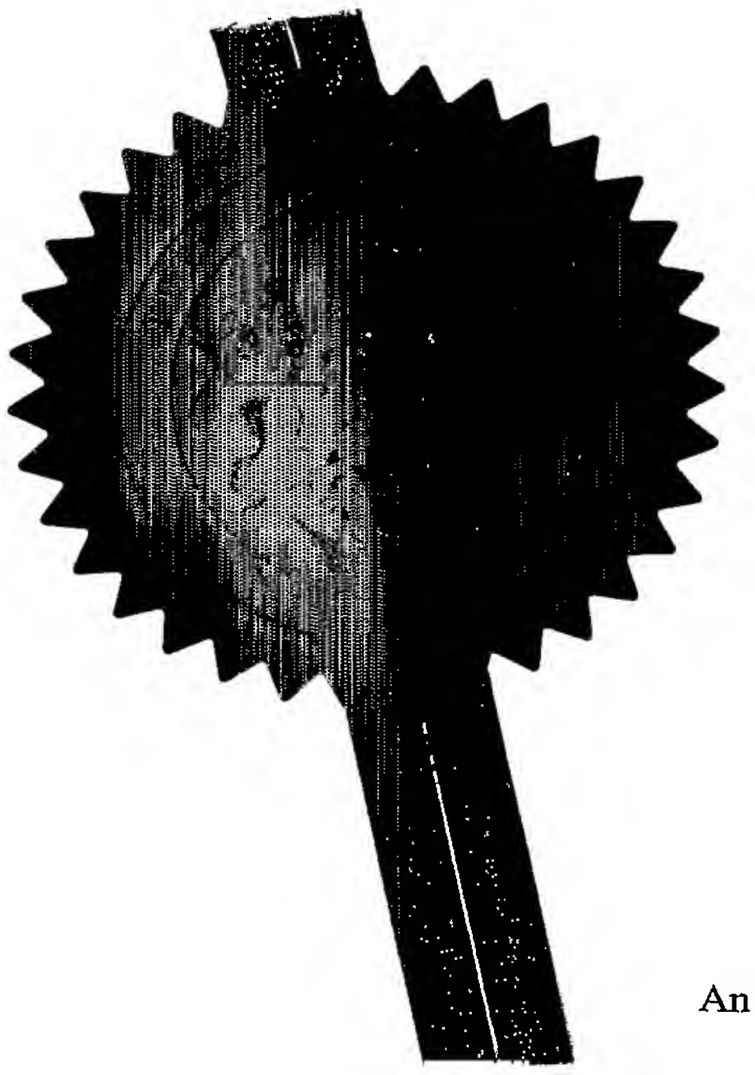

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3. Full name, address and postcode of the or of each applicant *(underline all surnames)* — NORCHIP AS
Industriveien 8,
N-3490 KLOKKARSTUA,
NORWAY

Patents ADP number *(if you know it)* — 81070062001

If the applicant is a corporate body, give the country/state of its incorporation — NORWAY

4. Title of the invention — IMPROVED DETECTION OF HUMAN PAPILLOMAVIRUS

5. Name of your agent *(if you have one)* — BOULT WADE TENNANT

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Signature: Bourne Nicole Tennent

Date: 26 February 2004

12. Name and daytime telephone number of person to contact in the United Kingdom: Nina L. White 020-7430-7500

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IMPROVED DETECTION OF HUMAN PAPILLOMAVIRUS

Field of the invention

The present invention relates to *in vitro* methods of screening human subjects for the presence of human papillomavirus which exhibits loss of regulation of E6/E7 mRNA expression and loss of replication. The methods are useful in the context of cervical cancer screening.

Background to the invention

Cervical carcinoma is one of the most common malignant diseases world-wide and is one of the leading causes of morbidity and mortality among women (Parkin DM, Pisani P, Ferlay J (1993) Int J Cancer 54: 594-606;Pisani P, Parkin DM, Ferlay J (1993) Int J Cancer 55: 891-903). 15,700 new cases of invasive cervical cancer were predicted in the United States in 1996, and the annual world-wide incidence is estimated to be 450,000 by the World Health Organization (1990). The annual incidence rate differs in different parts of the world, ranging from 7.6 per 100,000 in western Asia to 46.8 per 100,000 in southern Africa (Parkin et al., 1993 *ibid*).

The current conception of cervical carcinoma is that it is a multistage disease, often developing over a period of 10-25 years. Invasive squamous-cell carcinoma of the cervix is represented by penetration through the basal lamina and invading the stroma or epithelial lamina propria. The clinical course of cervical carcinoma shows considerable variation. Prognosis has been related to clinical stage, lymph node involvement, primary tumour mass, histology type, depth of invasion and lymphatic permeation (Delgado G, et al., (1990) Gynecol Oncol 38: 352-357). Some patients with less favourable tumour characteristics have a relatively good outcome, while others suffer a fatal outcome of an initially limited disease. This

shows a clear need for additional markers to further characterise newly diagnosed cervical carcinomas, in order to administer risk-adapted therapy (Ikenberg H, et al., Int. J. Cancer 59:322-6. 1994).

The epidemiology of cervical cancer has shown strong association with religious, marital and sexual patterns. Almost 100 case-control studies have examined the relationship between HPV and cervical neoplasia and almost all have found positive associations (IARC monographs, 1995). The association is strong, consistent and specific to a limited number of viral types (Munoz N, Bosch FX (1992) HPV and cervical neoplasia: review of case-control and cohort studies. IARC Sci Publ 251-261). Among the most informative studies, strong associations with HPV 16 DNA have been observed with remarkable consistency for invasive cancer and high-grade CIN lesions, ruling out the possibility that this association can be explained by chance, bias or confounding (IARC monographs, 1995). Indirect evidence suggested that HPV DNA detected in cancer cells is a good marker for the role of HPV infection earlier in the carcinogenesis. Dose-response relationship has been reported between increasing viral load and risk of cervical carcinoma (Munoz and Bosch, 1992 *ibid*). In some larger series up to 100% of the tumours were positive for HPV but the existence of virus-negative cervical carcinomas is still debatable (Meijer CJ, et al., (1992) Detection of human papillomavirus in cervical scrapes by the polymerase chain reaction in relation to cytology: possible implications for cervical cancer screening. IARC Sci Publ 271-281; Das BC, et al., (1993) Cancer 72: 147-153).

The most frequent HPV types found in squamous-cell cervical carcinomas are HPV 16 (41%-86%) and 18 (2%-22%). In addition HPV 31, 33, 35, 39, 45, 51, 52, 54, 56, 58, 59, 61, 66 and 68 are also found (IARC, monographs, 1995). In the HPV2000 International

conference in Barcelona HPV 16, 18, 31 and 45 were defined as high risk, while HPV 33, 35, 39, 51, 52, 56, 58, 59, 68 were defined as intermediate risk (Keerti V. Shah. P71). The 13 high risk plus intermediate risk HPVs are together often referred to as cancer-associated HPV types.

A number of studies have explored the potential role of HPV testing in cervical screening (see Cuzick et al. A systematic review of the role of human papillomavirus testing withing a cervical screening programme. Health Technol Assess 3:14. 1999).

Reid et al., (Reid R, et al., (1991) Am J Obstet Gynecol 164: 1461-1469) where the first to demonstrate a role for HPV testing in a screening context. This study was carried out on high-risk women from sexually transmitted disease clinics and specialist gynaecologists, and used a sensitive (low stringency) Southern blot hybridisation for HPV detection. A total of 1012 women were enrolled, and cervicography was also considered as a possible adjunct to cytology. Twenty-three CIN II/III lesions were found altogether, but only 12 were detected by cytology (sensitivity 52%, specificity 92%). HPV testing found 16 high-grade lesions.

Bauer et al. (Bauer HM, et al., (1991) JAMA 265: 472-477) report an early PCR-based study using MY09/11 primers (Manos M, et al.,(1990) Lancet 335: 734) in young women attending for routine smears (college students). They found a positive rate of 46% in 467 women, which was much higher than for dot blot assay (11%).

In a study using PCR with GP5/6 primers (Van Den Brule AJ, et al., (1990) J Clin Microbiol 28: 2739-2743) van der Brule et al. (Van Den Brule AJ, et al., (1991) Int J Cancer 48: 404-408) showed a very strong correlation of HPV positivity with cervical

neoplasia as assessed by cytology. In older women (aged 35-55 years) with negative cytology the HPV positivity rate was only 3.5%, and this was reduced to 1.5% if only types 16, 18, 31 and 33 were considered, while women with histological carcinoma in situ were all HPV-positive, and 90% had one of the four above types. Women with less severe cytological abnormalities had lower HPV positivity rates in a graded way, showing a clear trend.

Roda Housman et al. (Roda Housman AM, et al., (1994) Int J Cancer 56: 802-806) expanded these observations by looking at a further 1373 women with abnormal smears. This study also confirmed increasing positivity rate with increasing severity of smear results. They also noted that the level of HPV heterogeneity decreased from 22 types for low-grade smears to ten "high-risk" types for high grade smears. This paper did not include any cytologically negative women, nor was cytological disease confirmed histologically.

Cuzick et al. (Cuzick J, et al., (1992) Lancet 340: 112-113; Cuzick J, et al., (1994) Br J Cancer 69: 167-171) were the first to report that HPV testing provided useful information for the triage of cytological abnormalities detected during random screening. In a study of 133 women, referral for coloposcopy they found a positive predictive value of 42%, which was similar to that for moderate dyskaryosis. The results were most striking for HPV 16, where 39 of 42 HPV 16 positive women were found to have high-grade CIN on biopsy. This study pointed out the importance of assessing viral load and only considered high levels of high-risk types as positive.

Cox et al. (Cox JT, et al., (1995) Am J Obstet Gynecol 172: 946-954) demonstrated a role for HPV testing using the Hybrid Capture™ system (DIGENE Corporation, Gaithersburg, MD, USA) for triaging women

with borderline smears. This test was performed on 217 such women from a college referral service, and a sensitivity of 93% was found for CINII/III compared with 73% for repeat cytology. High viral load was found to further improve performance by reducing false positives. When 5 RLU was taken as a cut-off, a PPV of approximately 24% was found with no loss of sensitivity.

WO 91/08312 describes methods for determining the prognosis of individuals infected with HPV which comprise measuring the level of HPV activity by detecting transcripts of all or a portion of the E6 and/or E7 HPV genes in a sample and comparing the measurements of HPV activity with a previously established relationship between activity and risk of progression to serious cervical dysplasia or carcinoma.

WO 99/29890 describes methods for the assessment of HPV infection based on the measurement and analysis of gene expression levels. In particular, WO 99/29890 describes methods which are based on measuring the levels of expression of two or more HPV genes (e.g. HPV E6, E7, L1 and E2) and then comparing the ratio of expression of combinations of these genes to provide an indication of the stage of HPV-based disease in a patient.

The present inventors have previously determined that it is possible to make a clinically useful assessment of HPV-associated disease based only on a simple positive/negative determination of expression of E6 mRNA transcripts, with no requirement for accurate quantitative measurements of expression levels. This method is technically simple and, in a preferred embodiment, is amenable to automation in a mid-to-high throughput format. This method is described in detail in the applicant's published International application WO 03/57914.

The method described in WO 03/57914 is preferably carried out using the Pre-Tect HPV-Proofer™ kit, which is commercially available from Norchip AS. The HPV-Proofer assay provides three levels of information:
(1) Identification of mRNA from five specific different HPV-types (16, 18, 31, 33 and 45);
(2) Determination of presence of oncogene HPV E6/E7 mRNA; and
(3) Determination of presence of full length E6/E7 mRNA indicating dysregulation.

Each sample undergoes three duplex NASBA reactions, therefore six results are reported for each sample. Negative controls are included each time to monitor contamination. Positive controls are included for all HPV types to monitor reagent performance. Intrinsic cellular control U1A mRNA (cellular housekeeping gene) monitors entire test procedure to eliminate possible false negatives. It is not possible for the HPV-Proofer assay to detect HPV DNA. PreTect Analysis Software (PAS) for automated routine data analysis, interpretation and reporting.

The utility of the HPV-Proofer assay has been evaluated in at least 12 clinical studies.

The present inventors have now determined that the Pre-Tect HPV-Proofer™ assay does not detect HPV virions, even though it detects HPV-mRNA from HPV 16, 18, 31, 33 and 45. Rather, presence of E6/E7 transcript, as may be determined with the HPV-Proofer assay, is indicative of a loss of transcriptional regulation and loss of ability to replicate. It is impossible for the Pre-Tect HPV-Proofer to detect infectious virus HPV particles, since the virus cannot produce virions when transcriptional regulation is lost and the virus is integrated. The virus inside the cells detected as positive for E6/E7 expression using HPV-Proofer have left their normal life-cycles

and lost their regulation of either transcription control of the promoters from all the E6/E7 transcripts, or lost the splicing capability, leaving the E6/E7 transcript behind.

The inventors have also observed that the expression of E6/E7 mRNA transcripts in HPV-infected cells correlates with cellular changes characterised as the presence of enlarged cell nuclei, aneuploidy (typically more than 5 or 9 centromeres per cell) and also mitosis. Cells that are positive for E6/E7 expression (e.g. using the PreTect HPV-Proofer test) have something wrong, they exhibit cell abnormalities or have large maturated cell nuclei. These results also correlate with cytological and histological characterisation of cervical lesions. Cytologically or histologically defined low-grade lesions lacking cells with enlarged cell nuclei and with less than 9 centromeres do not give positive results for expression of E6/E7 mRNA expression with HPV-Proofer.

Therefore, the inventors have determined that expression of E6/E7 transcripts of human papillomavirus can be used as a molecular indicator of the presence of cellular abnormalities associated with the presence of a persistent infection with human papillomavirus. Detection of E6/E7 expression may therefore be used to distinguish between high and low grade cervical lesions. In particular, detection of E6/E7 expression can discriminate between histologically-defined CIN III samples without aneuploidic cells and those having aneuploidic cells. Detection of E6/E7 expression can also distinguish between histologically defined CIN III or CIN II (HSIL cases) cases that go into regress and those in which infection persists or progresses.

The present inventors have still further concluded that:

(1) Incidence of expression from E6/E7 oncogenes increases with the severity of the lesion.
(2) Detecting HPV oncogenic activity by assessment of E6/E7 expression in combination with typing promises to be a powerful predictor of high-grade lesions.
(3) The significant majority of cervical cancers (96%) contain at least one of the five main carcinogenic HPV-types (HPV 16, 18, 31, 33 and 45) [in Norwegian study population].
(4) Those who tested positive for E6/E7 expression with Pre-Tect HPV-Proofer™ were significantly more likely to maintain a persistent infection than those without positive HPV-Proofer results.

Accordingly, in a first aspect the invention provides an *in vitro* method of screening human subjects for the presence of human papillomavirus in at least one cell or tissue, wherein the human papillomavirus exhibits loss of regulation of E6/E7 mRNA expression and loss of replication, the method comprising detecting the presence of mRNA transcripts of the E6/E7 gene of a human papillomavirus in a test sample comprising mRNA derived from the cell or tissue, wherein the presence of E6/E7 mRNA transcripts in the sample is taken as an indication of the presence of human papilloma virus exhibiting loss of regulation of E6/E7 mRNA expression and loss of replication in the cell or tissue.

In a second aspect the invention provides an *in vitro* method of screening human subjects for the presence of cellular changes characterized by enlarged cell nuclei and cellular aneuploidy in at least one cell or tissue, which method comprises detecting the presence of mRNA transcripts of the E6/E7 gene of human papillomavirus in a test sample comprising mRNA derived from the cell or tissue, wherein the presence of E6/E7 mRNA transcripts in the sample is taken as an indication that the cell or tissue under test exhibits the cellular changes.

A positive screening result in the methods of the invention is indicated by detection of expression of E6/E7 mRNA transcripts. A positive result for E6/E7 mRNA expression indicates that the subject carries virus which exhibits loss of regulation of E6/E7 expression and is further indicative that the subject has abnormal cell changes.

The term "loss of E6/E7 regulation" as used herein means a loss of regulation of either transcription control of the promoters from all the E6/E7 transcripts, or a loss of the normal splicing capability in the E6/E7 open reading frames.

The term "abnormal cell changes" encompasses cell changes which are characteristic of more severe disease than low-grade cervical lesions or low squamous intraepithelial lesions, includes cell changes which are characteristic of disease of equal or greater severity than high-grade CIN (defined as a neoplastic expansion of transformed cells), CIN (cervical intraepithelial neoplasia) III, or high squamous intraepithelial neoplasia (HSIL), including lesions with multiploid DNA profile and "malignant" CIN lesions with increased mean DNA-index values, high percentage of DNA-aneuploidy and 2.5c Exceeding Rates (Hanselaar *et al.*, 1992, Anal Cell Pathol., 4:315-324; Rihet *et al.*, 1996, J. Clin Pathol 49:892-896; and McDermott *et al.*, 1997, Br. J. Obstet Gynaecol. 104:623-625).

Cervical Intraepithelial Neoplasia (abbreviated "CIN"), also called Cervical Dysplasia, is a cervical condition caused Human Papilloma Virus. CIN is classified as I, II or III depending on its severity. It is considered a pre-cancerous abnormality, but not an actual cancer. The mildest form, CIN I, usually goes away on its own, although rarely it can progress to cancer. The more severe forms, CIN II and CIN III,

most often stay the same or get worse with time. They can become a cancer, but almost never do if treated adequately.

HPV has been identified as a causative agent in development of cellular changes in the cervix, which may lead to the development of cervical carcinoma. These cellular changes are associated with constitutive or persistent expression of E6/E7 proteins from the HPV viral genome. Thus, it is possible to conclude that subjects in which expression of E6 mRNA can be detected, particularly those subjects who exhibit persistent E6 expression when assessed over a period of time, already manifest cellular changes in the cervix. These changes may have taken place in only a very few cells of the cervix, and may not be detectable by conventional cytology. Nevertheless, with the use of sensitive, specific and accurate methods for detection of E6/E7 mRNA it is possible to identify those subjects who already exhibit cellular changes in the cervix at a much earlier stage than would be possible using conventional cytological screening. This will allow earlier intervention with treatments aimed at preventing the development of cervical carcinoma.

As a result of HPV integration into the human genome or as a result of the "modification" in a modified episomal HPV genome, normal control of the viral E6/E7 oncogene transcription is lost (Durst et al., 1985, J Gen Virol, 66(Pt 7): 1515-1522; Pater and Pater, 1985 Virology 145:313-318; Schwarz *et al.*, 1985, Nature 314: 111-114; Park *et al.*, 1997, *ibid*). In contrast, in premalignant lesions and HPV-infected normal epithelium papillomaviruses predominate in "unmodified" episomal forms, hence oncogene (E6/E7) transcription may be absent or efficiently down-regulated (Johnson *et al.*, 1990, J Gen Virol, 71(Pt 7): 1473-1479; Falcinelli *et al.*, 1993, J Med Virol, 40: 261-265). Integration of human

papillomavirus type 16 DNA into the human genome is observed to lead to a more unstable cell activity/genome, and increased stability of E6 and E7 mRNAs (Jeon and Lambert, 1995, Proc Natl Acad Sci USA 92: 1654-1658). Thus HPV integration, typically found in cervical cancers but only infrequently found in CIN lesions (Carmody *et al.*, 1996, Mol Cell Probes, 10: 107-116), appears to be an important event in cervical carcinogenesis.

In a clinical context the performance of methods which rely on screening for expression of E6/E7 mRNA alone is critically dependent on the ability to score a negative result for E6/E7 mRNA expression with confidence. This again requires a detection technique which has maximal sensitivity, yet produces minimal false-negative results. In a preferred embodiment this is achieved by using a sensitive amplification and real-time detection technique to screen for the presence or absence of E6/E7 mRNA. The most preferred technique is real-time NASBA amplification using molecular beacons probes, as described by Leone et al., Nucleic Acids Research., 1998, Vol 26, 2150-2155. Due to the sensitivity of this technique the occurrence of false-negative results is minimised and a result of "negative E6/E7 expression" can be scored with greater confidence. This is extremely important if the assays are to be used in the context of a clinical screening program.

It is preferred to assay for expression of E6/E7 mRNA transcripts from any one or more of (and more preferably all of) HPV types 16, 18, 31, 33 and 45. In one embodiment the assay may detect only these HPV types. DNA from HPV types 16, 18, 31 and 33 has been detected in more than 96%% of cervical carcinoma samples in a Norwegian study population. Other studies have shown that E6 and E7 are almost invariably retained in cervical cancers, as their expression is likely to be necessary for conversion to

and maintenance of the malignant state (Choo *et al.*, 1987, J Med Virol 21:101-107; Durst *et al.*, 1995, Cancer Genet Cytogenet, 85: 105-112). In contrast to HPV detection systems which are based on detection of the undamaged genome or the L1 gene sequence, detection of HPV mRNA expressed from the E6/E7 area may detect more than 90% of the patients directly related to a risk of developing cervical carcinoma.

In the clinic, methods based on detection of E6/E7 mRNA may be used in post-screening, i.e. further analysis of individuals having a previous diagnosis of ASCUS, CIN 1 or Condyloma. The method may be used to select those with a high risk of developing cervical carcinoma from amongst the group of individuals having a previous diagnosis of ASCUS, CIN 1 or Condyloma. ASCUS, Condyloma and CIN I may be defined as more or less the same diagnosis due to very low reproducibility between different cytologists and different cytological departments. Östör (Int J. Gyn Path. 12:186-192. 1993) found that only around 1% of the CIN 1 cases may progress to cervical carcinoma. Thus, there is a genuine need for an efficient method of identifying the subset of individuals with ASCUS, Condyloma or CIN I who are at substantial risk of developing cervical carcinoma. One of HPV types 16, 18, 31 or 33 was detected in 87% of the cervical carcinoma cases study by Karlsen *et al.*, 1996. By inclusion of HPV 45, nearly 90% of the cervical carcinoma samples are found to be related to these five HPV types. Therefore, calculated from the data provided by Östör (Int J. Gyn Path. 12:186-192. 1993) more than 99.9% are detected cases with ASCUS, CIN I or condyloma are missed by our HPV-Proofer kit.

The high sensitivity and specificity of the present method means that it may find utility in primary screening, reflex-testing kit or routine diagnostics for detection of women with a high or very high risk of developing cervical carcinoma.

In the method of the invention "positive expression" of an mRNA is taken to mean expression above background. There is no absolute requirement for accurate quantitative determination of the level of E6/E7 mRNA expression.

In certain embodiments, the methods of the invention may comprise a quantitative determination of levels of mRNA expression. In a preferred embodiment in order to provide a clear distinction between "positive expression" and "negative expression" a determination of "positive expression" may require the presence of more than 50 copies of the relevant mRNA (per ml of sample or per total volume of sample), whereas a determination of "negative expression" may require the presence of less than 1 copy of the relevant mRNA (per ml of sample or per total volume of sample).

The methods of the invention will preferably involve screening for E6/E7 mRNA using a technique which is able to detect specifically E6/E7 mRNA from cancer-associated HPV types, more preferably "high risk" cancer-associated HPV types. In the most preferred embodiment the methods involve screening for E6/E7 mRNA using a technique which is able to detect E6 mRNA from HPV types 16, 18, 31 and 33, and preferably also 45. Most preferably, the method will specifically detect expression of E6/E7 mRNA from at least one of HPV types 16, 18, 31, 33, and preferably also 45, and most preferably all five types. However, women positive for positive for expression of E6/E7 from other types than 16, 18, 31, 33 and 45, e.g. 35, 39, 45, 52, 56, 58, 59, 66 and 68 may still manifest cellular abnormalities. Thus, the method may encompass screening for expression of E6/E7 mRNA from one or more of these HPV types, most preferably in addition to screening for E6/E7 mRNA from HPV types 16, 18, 31, 33 and 45. Certain HPV types exhibit a

marked geographical/population distribution. Therefore, it may be appropriate to include primers specific for an HPV type known to be prevalent in the population/geographical area under test, for example in addition to screening for HPV types 16, 18, 31, 33 and 45.

For the avoidance of doubt, unless otherwise stated the term "E6/E7 mRNA" as used herein encompasses naturally occurring mRNA transcripts which contain all or part of the E6 and/or E7 open reading frames. Preferred embodiments of the methods of the invention are based on detection of full length E6/E7 mRNA transcripts of some or all of the HPV types. In these embodiments presence of the full length E6/E7 mRNA is taken as a positive screening result.

The term "full length E6/E7 mRNA transcripts" excludes any of the naturally occurring splice variants, but encompasses bicistronic transcripts that encode functional E6 and E7 proteins. Four E6/E7 mRNA species have so far been described in cells infected with HPV 16, namely an unspliced E6 transcript and three spliced transcripts denoted E6*I, E6*II and E6*III (Smotkin D, et al., J Virol. 1989 Mar 63(3):1441-7; Smotkin D, Wettstein FO. Proc Natl Acad Sci USA. 1986 Jul 83(13):4680-4; Doorbar J. et al., Virology. 1990 Sep 178(1):254-62; Cornelissen MT, et al. J Gen Virol. 1990 May 71(Pt 5):1243-6; Johnson MA, et al. J Gen Virol. 1990 Jul 71(Pt 7):1473-9; Schneider-Maunoury S, et al. J Virol. 1987 Oct 61(10):3295-8; Sherman L, et al. Int J Cancer. 1992 Feb 50(3):356-64). All four transcripts are transcribed from a single promoter (p97) located just upstream of the second ATG of the E6 ORF. In the case of HPV 16, the term "full length E6/E7 transcripts" refers to transcripts which contain all or substantially all of the region from nucleotide (nt) 97 to nt 880 in the E6 ORF, inclusive of nt 97 and 880. Nucleotide positions are numbered according to

standard HPV nomenclature (see Human Papillomavirus Compendium OnLine, available via the internet or in paper form from HV Database, Mail Stop K710, Los Alamos National Laboratory, Los Alamos, NM 87545, USA).

In relation to HPV types other than HPV 16, "full length" E6/E7 transcripts may be taken to include transcripts which contain sequences homologous to the above-stated region of the HPV 16 E6/E7 transcript and to exclude E6 splice variants. Various sequence alignments of HPV types are publicly available via the Human Papillomavirus Compendium OnLine.

Specific detection of full length E6/E7 mRNA transcripts may be accomplished, for example, using primers or probes which are specific for the region which is present only in full length E6/E7 transcripts, not in splice variants.

Different HPV types exhibit different patterns of E6/E7 mRNA expression. Transcript maps for various HPV types, including HPV types 16 and 31, which may be used to assist in the design of probes or primers for detection of E6/E7 transcripts are publicly available via the Human Papillomavirus Compendium (as above).

Assay methodology

The disclosed screening methods may be carried out on a preparation of nucleic acid isolated from a clinical sample or biopsy containing cervical cells taken from the subject under test. Suitable samples which may be used as a source of nucleic acid include (but not exclusively) cervical swabs, cervical biopsies, cervical scrapings, samples removed with the use of brushes and tampons etc., skin biopsies/warts, also paraffin embedded tissues, and formalin or methanol fixed cells.

The preparation of nucleic acid to be screened using the disclosed methods must include mRNA, however it need not be a preparation of purified poly A+ mRNA and preparations of total RNA or crude preparations of total nucleic acid containing both RNA and genomic DNA, or even crude cell lysates are also suitable as starting material for a NASBA reaction. Essentially any technique known in the art for the isolation of a preparation of nucleic acid including mRNA may be used to isolate nucleic acid from a test sample. A preferred technique is the "Boom" isolation method described in US-A-5,234,809 and EP-B-0389,063. This method, which can be used to isolate a nucleic acid preparation containing both RNA and DNA, is based on the nucleic acid binding properties of silicon dioxide particles in the presence of the chaotropic agent guanidine thiocyanate (GuSCN).

The methods of the invention are based on assessment of active transcription of the HPV genome. The methods are not limited with respect to the precise technique used to detect mRNA expression. Many techniques for detection of specific mRNA sequences are known in the art and may be used in accordance with the invention. For example, specific mRNAs may be detected by hybridisation, amplification or sequencing techniques.

It is most preferred to detect mRNA expression by means of an amplification technique, most preferably an isothermal amplification such as NASBA, transcription-mediated amplification, signal-mediated amplification of RNA technology, isothermal solution phase amplification, etc. All of these methods are well known in the art More preferably mRNA expression is detected by an isothermal amplification in combination with real-time detection of the amplification product. The most preferred combination is amplification by NASBA, coupled with real-time

detection of the amplification product using molecular beacons technology, as described by Leone et al., Nucleic Acids Research, 1998, Vol 26, 2150-2155.

Methods for the detection of HPV in a test sample using the NASBA technique will generally comprise the following steps:

(a) assembling a reaction medium comprising suitable primer-pairs, an RNA directed DNA polymerase, a ribonuclease that hydrolyses the RNA strand of an RNA-DNA hybrid without hydrolysing single or double stranded RNA or DNA, an RNA polymerase that recognises said promoter, and ribonucleoside and deoxyribonucleoside triphosphates;

(b) incubating the reaction medium with a preparation of nucleic acid isolated from a test sample suspected of containing HPV under reaction conditions which permit a NASBA amplification reaction; and

(c) detecting and/or quantitatively measuring any HPV-specific product of the NASBA amplification reaction.

Detection of the specific product(s) of the NASBA reaction (i.e. sense and/or antisense copies of the target RNA) may be carried out in a number of different ways. In one approach the NASBA product(s) may be detected with the use of an HPV-specific hybridisation probe capable of specifically annealing to the NASBA product. The hybridisation probe may be attached to a revealing label, for example a fluorescent, luminescent, radioactive or chemiluminescent compound or an enzyme label or any other type of label known to those of ordinary skill in the art. The precise nature of the label is not critical, but it should be capable of producing a signal detectable by external means, either by itself or in conjunction with one or more additional substances (e.g. the substrate for an enzyme).

A preferred detection method is so-called "real-time NASBA" which allows continuous monitoring of the formation of the product of the NASBA reaction over the course of the reaction. In a preferred embodiment this may be achieved using a "molecular beacons" probe comprising an HPV-specific sequence capable of annealing to the NASBA product, a stem-duplex forming oligonucleotide sequence and a pair of fluorescer/quencher moieties, as known in the art and described herein. If the molecular beacons probe is added to the reaction mixture prior to amplification it may be possible to monitor the formation of the NASBA product in real-time (Leone *et al.*, Nucleic Acids Research, 1998, Vol 26, 2150-2155). Reagent kits and instrumentation for performing real-time NASBA detection are available commercially (e.g. NucliSens™ EasyQ system, from Organon Teknika).

In a further approach, the molecular beacons technology may be incorporated into the primer 2 oligonucleotide allowing real-time monitoring of the NASBA reaction without the need for a separate hybridisation probe.

In a still further approach the products of the NASBA reaction may be monitored using a generic labelled detection probe which hybridises to a nucleotide sequence in the 5' terminus of the primer 2 oligonucleotide. This is equivalent to the "NucliSens™" detection system supplied by Organon Teknika. In this system specificity for NASBA products derived from the target HPV mRNA may be conferred by using HPV-specific capture probes comprising probe oligonucleotides as described herein attached to a solid support such as a magnetic microbead. Most preferably the generic labelled detection probe is the ECL™ detection probe supplied by Organon Teknika. NASBA amplicons are hybridized to the HPV-specific capture probes and the generic ECL probe (via a complementary sequence on primer 2).

Following hybridization the bead/amplicon/ECL probe complexes may be captured at the magnet electrode of an automatic ECL reader (e.g. the NucliSens™ reader supplied by Organon Teknika). Subsequently, a voltage pulse triggers the ECL™ reaction.

Preferred embodiments of the method rely on amplification of E6/E7 mRNA from at least the major cancer-associated HPV types 16, 18, 31 and 33, and preferably also HPV 45. There are several different ways in which this can be achieved.

In one embodiment, separate primer-pairs specific for each of HPV types 16, 18, 31 and 33, and preferably also HPV 45 may be used to amplify transcripts from each HPV type individually. Alternatively, mixtures of two or more primer-pairs in a single container may be used to enable multiplexing of the amplification reactions.

In a further embodiment, a single primer-pair capable of amplifying a region of the E6/E7 gene from HPV types 16, 18, 31 and 33, and preferably also HPV 45 may be used, which thus enables amplification of all four (preferably five) types in a single amplification reaction. This could, for example, be achieved with the use of a pair of degenerate primers or by selection of a region of the E6/E7 mRNA which is highly conserved across HPV types.

The E6/E7 primer-pair may correspond to any region of the E6/E7 mRNA, an may enable amplification of all or part of the E6 open reading frame and/or the E7 open reading frame. Preferably it will enable amplification of full length transcripts.

In a further approach, specificity for multiple HPV types may be achieved with the use of degenerate oligonucleotide primers or complex mixtures of polynucleotides which exhibit minor sequence

variations, preferably corresponding to sites of sequence variation between HPV genotypes. The rationale behind the use of such degenerate primers or mixtures is that the mixture may contain at least one primer-pair capable of detecting each HPV type.

In a still further approach specificity for multiple HPV types may be achieved by incorporating into the primers one or more inosine nucleotides, preferably at sites of sequence variation between HPV genotypes.

Lists of suitable primers and probes which may be used for the detection of E6/E7 mRNA from various HPV types may be found in WO 03/057914 and in WO 03/057927.

The method of the invention is preferably carried out using the Pre-Tect HPV-Proofer™ assay and kit. However, it is to be understood that the invention is not limited to the use of this specific assay.

The method of the present invention will score negative for real histological negative and representative samples from the whole or parts of cervix, corpus or/and the cervical canal. This provides the outstanding specificity that makes the PreTect HPV-Proofer™ one of the most promising primary screening methods ever developed.

The specificity of using PreTect HPV-Proofer™ alone for diagnostics of women at risk of developing cervical carcinoma has been proved to be independent of age and works with more than three times higher specificity than a commercial DNA-based assay alone.

Detection of E6/E7 transcripts has the potential to identify which high-risk infections may persist without having to perform repeat testing. Incidence of expression from E6/E7 oncogenes increases with the

severity of the lesion. Detecting HPV oncogenic activity in combination with typing promises to be a powerful predictor of high-grade lesions.

The invention will be further understood with reference to the following experimental examples and figures.

Pre-Tect HPV Proofer™ Assay

For all experimental examples which refer to the use of the Pre-Tect HPV Proofer™ kit and assay, the assay was performed using the commercially available kit according to the supplied instructions. Further information concerning the operation of the assay for real-time detection of HPV E6/E7 mRNA may be found in WO 03/057914, the entire contents of which are incorporated herein by reference.

Summary of clinical data

The attached experimental section summarises clinical data obtained using the Pre-Tect HPV-Proofer™ assay and kit.

PreTect®

HPV-Proofer

Routine assay - Total quality control

Providing three levels of information:

1. Identification of mRNA from five specific different HPV-types (16, 18, 31, 33 and 45)
2. Presence of oncogene HPV E6/E7 mRNA
3. Presence of full length E6/E7 mRNA indicating dysregulation

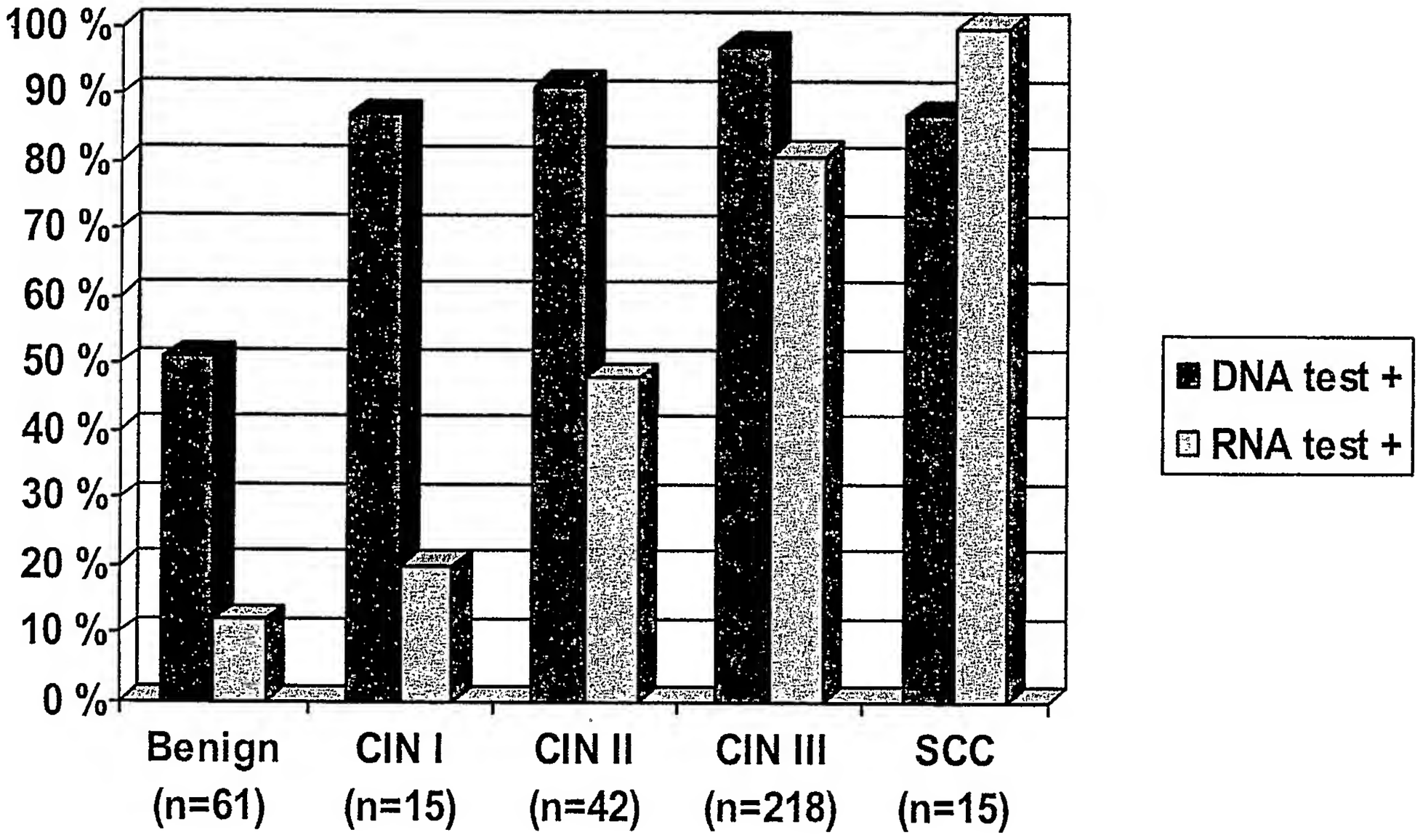
HPV-Proofer does not detect HPV virions!
No age discrimination (not only ">30 years")

- Each sample undergoes three duplex NASBA reactions
- Six results are reported for each sample
- Negative controls are included each time to monitor contamination
- Positive controls are included for all HPV types to monitor reagent performance
- Intrinsic cellular control U1A mRNA (cellular housekeeping gene) monitors entire test procedure to eliminate possible false negatives
- It is not possible for the HPV-Proofer to detect DNA
- PreTect Analysis Software (PAS) for automated routine data analysis, interpretation and reporting

Evaluation of HCII and HPV-Proofer

HPV testing related to histology (n=351)

HCII and HPV-Proofer was done on the same smear samples at the same time



Conclusions:

- "The RNA test revealed a higher prognostic value and higher specificity than the DNA test"
- HPV-Proofer detected all SCC vs. 87% for the DNA test
- The DNA test does not discriminate between high and low-grade lesions
- No age discrimination (not only ">30 years")

E6 and E7 mRNA expression in 4136 cervical smears

	Normal	ASCUS	Condyloma	CIN I	CIN II	CIN III	SCC	Total
HPV mRNA	95 2%	12 21%	6 32%	0 0%	2 40%	9 75%	1 100%	126 3%
HPV type-spec. PCR	144 4%	14 25%	9 47%	1 100%	2 40%	9 75%	1 100%	184 4%
HPV consensus DNA	368 9%	27 47%	14 74%	1 100%	2 40%	10 83%	1 100%	429 10%
Cytology	3970	57	19	1	5	12	1	4136*

- The combo:100% NPV, 100% sensitivity and 97% specificity
- Few samples with “normal” or “ASCUS” cytology had abnormal nucleus determined by E6/E7 full-length mRNA transcription
- > 50% of normal cytological samples, positive for HPV16/18 DNA, were E6/E7 mRNA negative

Follow up (2 years) study in Edinburgh (n=3445)

- Follow-up study of 54 cytological normal cases all positive for L1 HPV DNA and typed with line-blot. 28 was HPV-Proofer positive at baseline and 26 was negative
- HPV 16 DNA was the only HPV type without E6/E7 mRNA at baseline that manage to establish transcription of full-length mRNA after follow-up
- "Detection of mRNA E6/E7 transcripts identified which infections were more likely to persist, …."
  (Twice as high relative specificity for mRNA as DNA)
- "Detection of E6/E7 transcripts has the potential to identify which high-risk infections may persist without having to perform repeat testing"

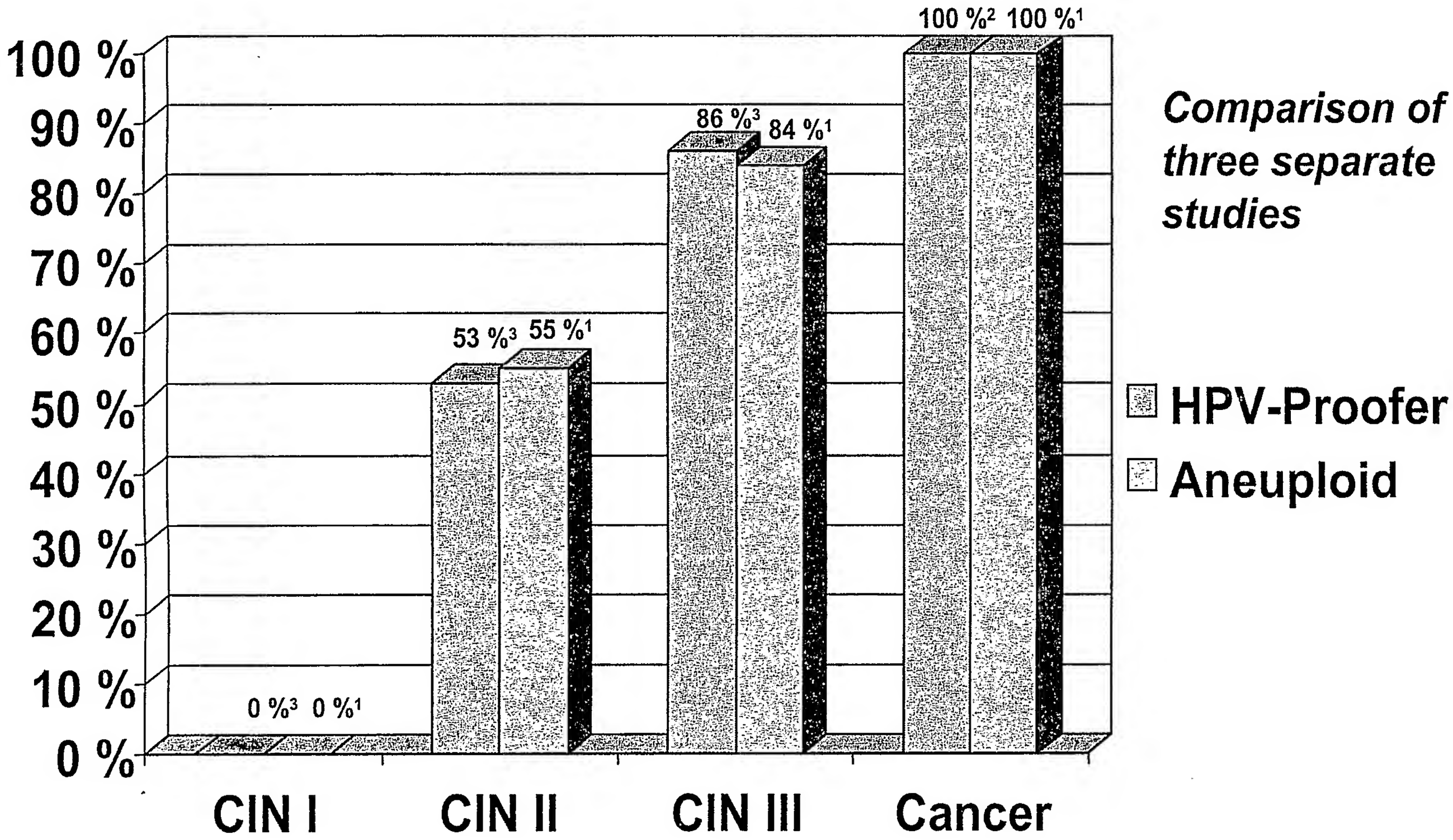
CIN III study (n=190)

- Only 37 biopsies (randomly collected from the cone) from 190 original cytological CIN III positive women was confirmed to be histological CIN III
- Histological CIN I (n=14) did not have active oncogenes from the carcinogenic HPV-types (16, 18, 31, 33 and 45)
- 34/36 CIN III biopsies, positive for HPV DNA from these five HPV-types, express E6/E7 mRNA

Cancer study (n=204)

- All samples evaluated were biopsies
- 89% of the samples had oncogenic expression from only five HPV-types (type specific PCR: 88%, consensus PCR: 92%)
- Compared to consensus PCR, the coverage rate of HPV-Proofer was 96,8% (North-America and Europe: >96% Clifford et al., 2003)

Rate of oncogene expression and DNA ploidy aberrations in biopsies



Comparison of three separate studies

Conclusions

1. Incidence of expression from E6/E7 oncogenes increases with the severity of the lesion

2. Detecting HPV oncogenic activity in combination with typing promises to be a powerful predictor of high-grade lesions

3. The significant majority of cancers (96%) contain at least one of the five main carcinogenic HPV-types (HPV 16, 18, 31, 33 and 45) [Norway]

4. Those who tested positive with HPV-Proofer were significantly more likely to maintain a persistent infection than those without positive HPV-Proofer results

Comparison between two studies

1. Head to head comparison of two different commercial methods in Norway.
   - Independent blindly study by the Norwegian Radium Hospital, Ullevål hospital, ST Olavs Hospital and the Norwegian Cancer Registry
   - 339 cytological normal and 269 cytological positive women was collected from an around 8000 outpatient women in Norway. Nineteen cervical smears from women with cervical carcinoma was added to the study from the Norwegian Radium Hospital.
   - Biopsies were taken if the HPV test was positive or cytology revealed HSIL. 383 histological diagnosis was performed.
   - 107 high-grade lesions (out of 271 high-grade) was not discovered with cytology
2. Head to head comparison of two different commercial methods in Sweden
   - Independent blindly study by the Central Hospital in Karlstad and the Karolinska University Hospital in Stockholm.
   - 240 CIN I and ASCUS was included from around 15000 outpatient women in Sweden
   - LEEP biopsies were taken in the HCII test was positive or cytology was positive. 118 LEEP biopsies were collected.
   - 29 high-grade lesions (out of 36) were not discovered with cytology three month before LEEP conization.

Results from this comparison

- **The study in Norway:**
  - 51% of the histological normal biopsies (n=61) were positive with HCII
  - 12% of the histological normal biopsies (n=61) were positive with HPV-Proofer
  - 90% of the smears collected from cervical carcinoma samples (n=20) were positive with HCII
  - 100% of the smears collected from cervical carcinoma samples (n=20) were positive with HPV-Proofer.
- **The study in Sweden:**
  - 73% of the histological normal LEEP biopsies (n=18) were positive with HCII.
  - 73% of the histological normal LEEP biopsies (n=18) were positive with cytology
  - 0% of the histological normal LEEP biopsies (n=18) were positive with HPV-Proofer.

Prevalens, specificity and sensitivity in four large studies

	Histology and Cytology	Molecular biology	Sensitivity	Specificity	Relative PPV	Relative NPV
Validation study (> 30 years)	339 cyt norm. 288 cyt pos. 61 hist norm. 322 hist pos.	HPV-Proofer	76%	81%		
		HCII	93%	40%		
Validation study (< 30 years)		HPV-Proofer	82%	70%		
		HCII	98%	20%		
Oslo study (n=4136)	3970 cyt norm. 95 cyt pos. 25 hist pos.	HPV-Proofer	86%	97%	9%	100%
		PCR consensus	93%	90%	3%	100%
Edinburgh study (follow up) (n=3445)	3445 cyt norm. 54 PCR pos.	HPV-Proofer	100%	78%	12%	100%
		PCR consensus	100%	40%	27%	100%
Karlstad study (follow up) (n= 15000)	127 cyt norm. 90 cyt pos. 18 hist norm. 100 hist pos.	HPV-Proofer	81%	85%	46%	95%
		HCII	91%	51%	31%	96%

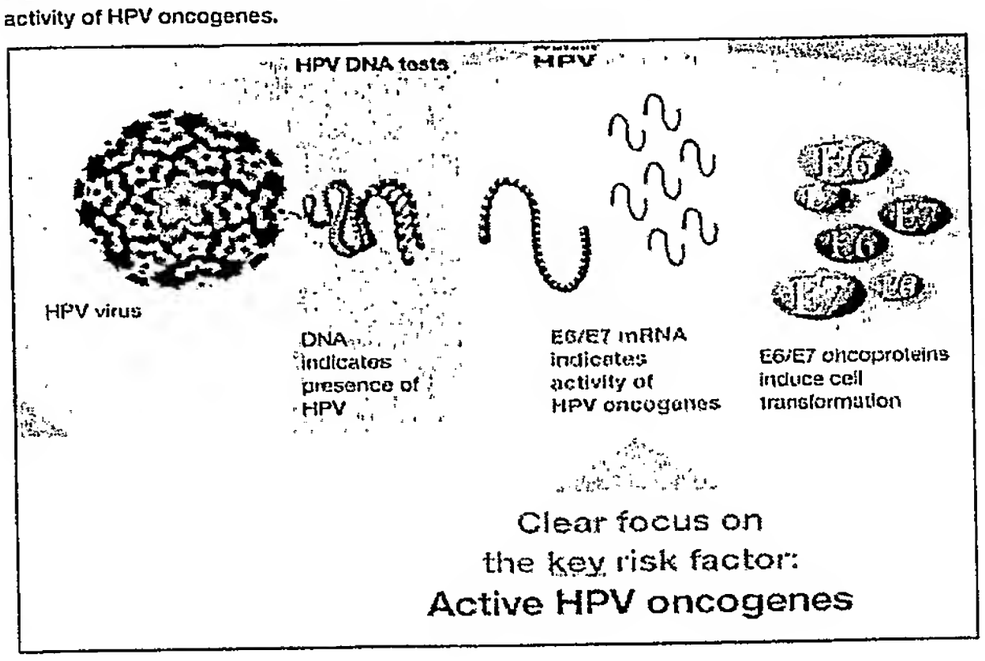
The potential of RNA as a target for molecular diagnostics in cervical carcinoma screening.

Karlsen F. NorChip AS, Klokkarstua, Norway

Introduction

Microarray technology and varied multiplex amplification methods (e.g. PreTect technology) have shown that RNA is valid as a target for routine molecular diagnostics and for future point-of-care testing. Detection of human papillomavirus targets may lend more accuracy and objectivity to the diagnostic process but consensus DNA-HPV detection methods have to be replaced with type-specific PCR and/or HPV RNA based detection methods, in order to increase the clinical specificity and positive predictive value (Table 2).
The association between E6-E7 gene products and cervical carcinogenesis may be the strongest ever-discovered including complex interactions with various cellular protein complexes, cell senescence and severe chromosomal instability (Figure 2) (von Knebel et al 2002). The main challenge using protein as a target for routine diagnostics has been low sensitivity, reproducibility and specificity. However, RNA and not DNA as a target for routine diagnostics may give the information of clinical activity, regulation or processes in addition to higher or equal sensitivity, reproducibility and specificity compared to DNA as target (Table 2). For more than 10 years ago new methods of isolation, purification and stabilization of mRNA was developed for routine diagnostics making the RNA very much suited as a marker for development of new diagnostics methods and even drugs. Available evidence indicated great potential for HPV testing within the cervical screening program. It offers the possibilities of greater sensitivity, reduced follow-up of low grade cytological abnormalities and treated lesions, increased screening intervals, and overall cost reduction (Cuzick 2002). An attractive screening option is to use HPV RNA and typing of a limited number of HPV types as the primary test and reserve cytology for triage to determine management.

Fig 1: HPV-Proofer detect E6/E7 mRNA from the five carcinogenic HPV types indicating activity of HPV oncogenes.



HPV-Proofer correlated with other methods

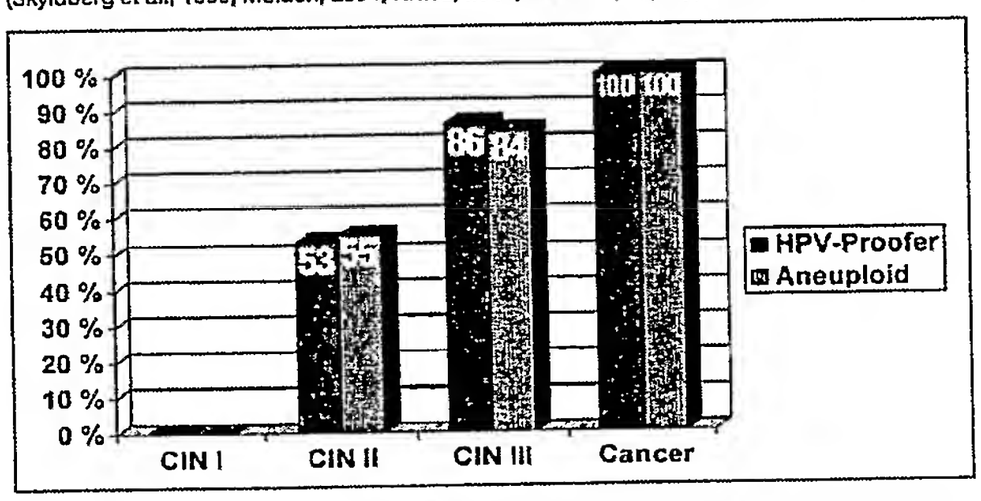
The HPV-Proofer method was developed to perform typing of HPV types 16, 18, 31, 33 and 45 with concurrent detection of expression markers (bicistronic mRNA of known oncogenes E6 and E7) in order to detect loss of viral and cell cycles regulation presented as a major cell nuclei abnormality. HPV-Proofer was evaluated against the following methods in the below listed clinical studies.

- Type-specific PCR
- Quantitative real-time PCR
- Consensus PCR
- HCII
- In situ hybridization
- Line-blot PCR
- NASBA with end-point ECL detection
- Singleplex real-time NASBA
- Sequencing analysis

Clinical studies

1. E6 and E7 mRNA expression in 4136 clinical samples (Molden, 2004 Poster number)
2. Development and verification of HPV-Proofer (Molden, 2003, submitted 2004)
3. High risk infections without oncogene expression in cervical smears (Molden 2004, Poster)
4. Cancer study on 204 squamous carcinoma samples (Lie, 2004, in prep 2004)
6. Follow up study in Edinburgh (Cuschieri, JMV, in press 2004, Poster)
7. Validation study in Oslo (Lie, 2004, Poster and Oral presentation)
8. Karlstad follow-up study in Sweden (Bjerre, 2003-2004, Poster)
9. Out-patience population study in DR Congo (Hovland, 2004)
10. High risk oncogene expression in progressive lesions (Nygaard, 2004, Poster and Oral pres.)
11. Evaluation of the main transcripts in clinical samples (Molden, in prep 2004)
12. Evaluation of the stability of mRNA from HPV 18 and 45 (Kraus, in prep 2004)

Fig 2:
Rate of mRNA expression in two clinical studies and DNA ploidy aberrations in biopsies (Skyldberg et al., 1999, Molden, 2004, Kraus, 2004, Lie et al., in prep).



This is two of our studies in blue compared with cytometry results done by Barbro Skyldberg at Karolinska Institutes

Table 1: mRNA and DNA analysis in 4136 cervical smears collected in a screening population in Norway.

	Normal	ASCUS	Condyloma	CIN I	CIN II	CIN III	SCC	Total
HPV mRNA	95 2%	12 21%	6 32%	0 0%	2 40%	9 75%	1 100%	126 3%
HPV DNA	368 9%	27 47%	14 74%	1 100%	2 40%	10 83%	1 100%	429 10%
Cytology	3970	57	19	1	5	12	1	4136*

*Results from groups unsatisfactory (64), AGUS (5) and ASC-H (2) not shown

Conclusion

1. Incidence of HPV oncogene expression increases with the severity of the lesion. **2.** Improved clinical specificity for HPV RNA compared to HPV DNA in "normal" and "ASCUS" samples. **3.** Detecting HPV oncogenic activity in combination with typing promises to be a powerful predictor of high-grade lesions. **4.** All of the cytological cell samples collected from women with cervical carcinoma were positive with HPV-Proofer (n=20). **5.** The significant majority of cancers (>96%) contain at least one of the five main carcinogenic HPV-types (HPV 16, 18, 31, 33 and 45). **6.** Those who tested positive with HPV-Proofer were significantly more likely to maintain a persistent infection than those without positive HPV-Proofer results. **7.** HPV-Proofer in combination with cytology gives the maximum security for physicians and patients (~100% sensitivity/specificity).

Table 2:
Prevalence, sensitivity and specificity in four large clinical studies

	Histology and Cytology	Molecular biology	Sensitivity	Specificity	Relative PPV	Relative NPV
Validation study (>30 years)	339 cyt norm. 288 cyt pos. 61 hist norm. 322 hist pos.	HPV-Proofer	76%	81%		
		HCII	93%	40%		
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		HCII	91%	51%	31%	96%

References
von Knebel et al., Eur J Cancer 2002, 38: 2229-42; Clifford et al., BJC 2003, 88:53-73.

DNA versus RNA based methods for HPV testing in Norway. Evaluation of Hybrid Capture II and PreTect HPV-Proofer, a validation study.

Lie AK[1], Risberg B[1], Sandstad B[2], Delabie J[1], Rimala R[3], Hagen B[4], Onsrud M[5], Thoresen S[6]
Department of Pathology (1) and Clinical Research (2), The Norwegian Radium Hospital, Oslo, Norway (1), Laboratory of Pathology, Oslo (3), Gynecologic Department, St. Olavs University Hospital, Trondheim (4), Gynecologic Department, Ullevål University Hospital, Oslo (5), Institute of Population-based Cancer Research, Oslo, Norway

Background

The specificity as well as the positive predictive value of HPV DNA testing in screening is low, because most high-risk HPV infections are transient. Expression of E6/E7 oncogenes are required for the development and maintenance of a malignant phenotype. HPV RNA testing can be used for risk evaluation and may have a superior positive predictive value in screening.

Aims

To validate two commercially available assays for HPV testing in order to investigate the prevalence of high-risk HPV infections in women with negative and positive cytology. To evaluate the outcome of DNA-based and RNA-based testing compared to cytology and histology.

Material and Methods

The study population was selected from outpatient departments and gynecologists in private practice. Included in this study were 628 women with median age 40 years (range, 19-85). A conventional Pap smear was taken first, and the remaining material was transferred to a PreservCyt™ vial (Cytyc Corporation). Testing for high-risk HPV DNA (type 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, and 68) was performed with the Hybrid Capture II assay (Digene Corporation) and individual identification of E6/E7 mRNA transcripts from HPV 16, 18, 31, 33, and 45 with the Pre Tect HPV-Proofer assay (NorChip AS), a real-time NASBA technique. Biopsies were taken when HPV test was positive or cytology revealed HSIL. Histology was regarded as the "gold standard".

Results

Concordance between cytology and histology were found in 53% of cases. High-grade histology (CIN 2+) was detected in 61% of the women with benign or low-grade cytology. Kappa value was 0.31. Different outcomes of the two tests were present in 17% (109/628) of cases (table 1). The HPV results related to cytology and histology diagnoses are shown in figure 1 and 2. Both HPV tests showed significant association with grade of the lesions ($p<0.001$). The DNA test was more often positive in benign and low-grade lesions. The DNA test revealed higher sensitivity but lower specificity compared to the RNA test (Table 2).

Conclusion

The RNA test revealed a higher prognostic value and higher specificity than the DNA test. Larger scale studies are necessary to evaluate the predictive values of these tests in the Norwegian screening program. National monitoring of HPV testing should be obligate and [illegible] cytology/histology results.

Table 1: HPV testing related to histology in cases with different outcome of the two HPV tests.

	Histology		
HPV test	< CIN 2	≥ CIN 2	Total
DNA +/ RNA-	40	59	99
RNA +/ DNA-	1	9	10
Total	41	68	109

Table 2: The performance of the HPV testing for detection of histological confirmed CIN 2+

	Sensitivity (%)		Specificity (%)	
	DNA	RNA	DNA	RNA
Age				
< 30 years (n=102)	98	82	20	70
≥ 30 years (n=281)	93	76	40	81
Cytology*				
normal (n=105)	89	62	79	87
low-grade (n=73)	96	72	22	72
high-grade (182)	96	83	67	50

* Twenty-three PAP smears with an unsatisfactory diagnosis were excluded from this analysis.

Fig 1: Cytological diagnoses related to HPV test.

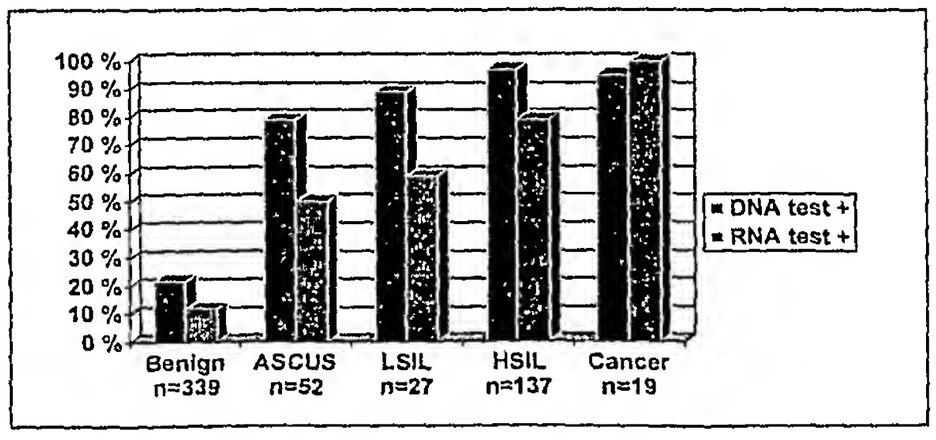
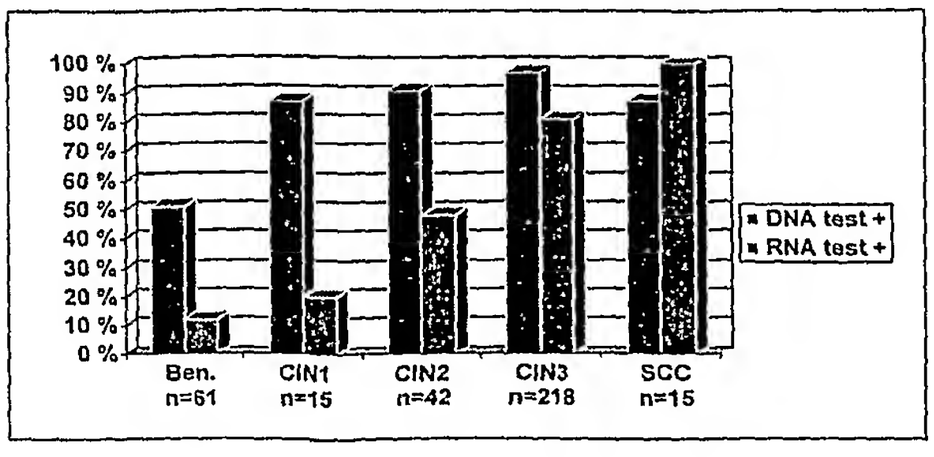


Fig 2: Histological diagnoses of squamous lesions related to HPV test.



HPV detection as follow-up of low-grade lesions in the Swedish gynaecological screening program. Comparison between Hybrid Capture II and the RNA-based PreTect HPV-Proofer Assay.

Bjerre P[1], Hagmar B[2], Dillner L [3], Edvardsson H[1], Mellinger I[4], Karlsson E[5], Karlsen F[6], Skomedal H[6] and Andersson-Ellstrøm A[1].
(1) Central Hospital, Karlstad, Sweden (2) The Norwegian National Hospital, Oslo, Norway (3) Karolinska Hospital, Stockholm, Sweden (4) Gynaecological polyclinics, Arvika, Sweden
(5) Gynaecological polyclincs, Saffle, Sweden (6) NorChip, Klokkarstua, Norway

Introduction

In Sweden approximately 40 000 cytology cases pr. year show aberrations which needs follow-up. Most cases regress spontaneously but some progress if not treated. There is also a problem of low sensitivity for cytology in the follow-up predure. In detection of pre- cancerous lesions, both specificity and sensitivity has been found to improve drastically when HPV testing is performed after detection of cytological ASCUS or CIN I.

Objective

The main objective was to evaluate the respective roles of HPV RNA and DNA tests in relation to cytology and histology in the Swedish screening program. Another important objective was to estimate the risk of missing CIN II+ in women with CIN I or ASCUS but negative with either HPV RNA or DNA tests.
The results will be of use for the follow-up routines and treatment strategies in Sweden.

Materials and Methods

Our material stems from 15000 women following the normal screening program in the central part of Sweden. All women positive for ASCUS or CIN I with cytology were selected for further studies. All the cytological or histological material was re-evaluated blindly by an experienced pathologist.
The samples positive for ASCUS and CIN I (N=240) were evaluated with PreTect HPV-Proofer (N=240), and a randomised selection of samples was tested by Hybrid Capture II (HCII) and cytology (N=127) and cytology alone (N=112) after 4 months. They were compared with histology from LEEP biopsies (N=126) after 7 months and with PreTect HPV-Proofer (N=240), HCII and cytology after 12 months (Table 1).
All samples with ASCUS and CIN I were tested for mRNA. Coloposcopy directed LEEP biopsies (N=126) were taken as a part of the follow-up for all women with an abnormal cytology diagnosis and/or positive HPV DNA test (after 4 months). HPV DNA was detected using the HCII assay (Digene, Gatesburg, MD, USA). Identification and individual typing of E6/E7 mRNA transcripts from HPV 16, 18, 31, 33, and 45 was carried out using the PreTect HPV-Proofer assay (NorChip AS, Klokkarstua, Norway).

Results

The results of HPV tests have been compared with cytology 4 and 12 months after and with histology diagnosis 7 months after positive cytology diagnosis (Table 1). Frequency and distribution of HPV types is presented in table 3. Concordance between cytology and histology was found in 19% of cases (Table 2). Cytology and the DNA test were considerably more often positive in benign and low-grade lesions by histology than the RNA test (Fig. 1). With histology as the "golden standard", the RNA test revealed a higher positive predictive value, and higher specificity (46% and 85,3% respectively) than the DNA test (31% and 51% respectively). However, the DNA test revealed a higher sensitivity (91%) than the RNA based test (81%). 19% of the cases treated with LEEP conization showed aberrant cytology 5 months after treatment (Table 1), 0.5% were found to be CIN II+. HPV DNA was detected in 24% and HPV RNA was detected in 6% of these cases (Table 1).

Discussion and conclusion

The higher postive predictive value and higher specificity of the RNA based method compared with the DNA based method may be explained by the fact that expression of the E6/E7 oncogenes is required for development and maintainance of the malignant phenotype. Frequency of RNA detection is somewhat higher in histological CIN II and lower in histological CIN III than previously seen in a Norwegian study (Kraus *et al.*, in press, JMV 2004). This may be due to differences in evaluation of histolgy between countries. The risk of missing CIN II+ in women with CIN I or ASCUS, but negative with either HPV RNA or DNA tests was extremely low (0.2%), confirming the added value of HPV testing in cytological ASCUS or CIN I.

Table 1: Overall results

	0 month	4 month	7 month	12 month
Cytology	240/240 (100%)	93/217 (43%)	Not analysed	30/160 (19%)
HCII	Not analysed	64/113 (57%)	Not analysed	41/169 (24%)
PreTect HPV-Proofer	Not analysed	56/240 (23%)	Not analysed	14/231 (6%)
Histology	Not analysed	Not analysed	100/118 (85%)	Not analysed

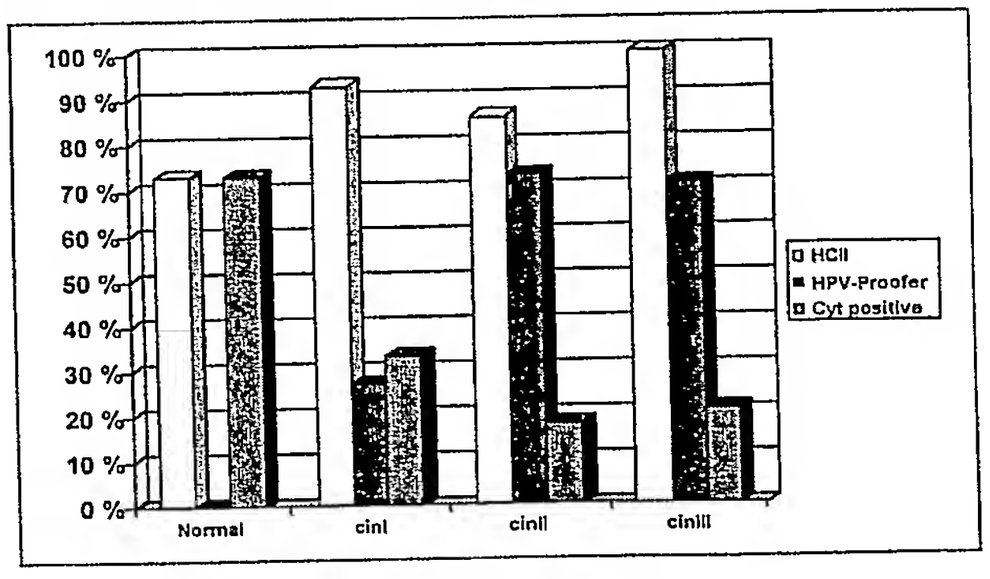
Table 2: Histology *versus* Cytology 4 months after positive diagnosis.

		Histology						
		Normal	Other	CIN I	CIN II	CIN III	Not analysed	TOTAL
Cytology	Normal	4	4	9	4	2	104	127
	Inadequate	1	1	1	2	0	4	9
	ASCUS/ ASCUS H	9	2	22	9	3/2	4	49/2
	CIN I	4	2	17	3	1	0	27
	CIN II	0	0	1	4	3	0	8
	CIN III	0	0	1	0	3	0	4
	Not analysed	0	0	0	0	0	14	14
	TOTAL	18	9	51	22	14	126	240

Table 3: Histological results *versus* HPV DNA, RNA and cytological results

Cytological diagnosis	ASCUS	CIN I	CIN II	CIN III/ASCUS-H
HPV 16	6	4	4	2
HPV 18	2	1	0	3
HPV 31	0	2	1	1
HPV 33	3	2	1	2
HPV 45	3	0	0	1
HPV-Proofer Total	14/49 (29%)	9/27 (33%)	5/8 (63%)	6/6 (100%)
HCII	20/26 (77%)	11/15 (73%)	3/3 (100%)	2/2 (100%)
Histology (CIN II+) only cyt.	3/23 (13%)	3/12 (25%)	4/5 (80%)	3/4 (75%)
Histology (CIN II+) cyt. & HCII	7/20 (35%)	1/11 (9%)	3/3 (100%)	2/2 (100%)
Histology (CIN II+) all samples	12/45 (27%)	4/27 (14%)	7/8 (88%)	5/6 (83%)

Fig 1: Cytology *versus* HPV test (4 months) and histology (7 months)



High-risk HPV infections without oncogene expression in women younger than 30 years of age

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Objective:

Human papillomavirus (HPV) is a common virus infection among women, particularly in younger age groups, although most infections are transient and asymptomatic. In the Scandinavian countries, the HPV prevalence in the women population above 30 years of age varies between 5 and 15% and the HPV prevalence in younger women may be as high as 30-40%. Also, 70 - 80% of the sexually active women will, at some point in their lifetime, acquire an HPV infection. However, the majority of the infections will spontaneously clear out, and only a small proportion will persist and give rise to cervical intraepithelial neoplasia (CIN). Expression from the E6/E7 oncogenes HPV is the main reason for cell changes and cell abnormalities in the cervix.
The aim of this study was to compare the detection of E6/E7 transcripts and the detection of HPV DNA in women younger than 30 years of age.

Material and Methods:

A total of 282 cervical samples from women younger than 30 years of age (mean age 26.9) were tested. RNA and DNA were extracted using the NucliSens Extractor and E6/E7 mRNA expression from the carcinogenic HPV types 16, 18, 31, 33, and 45 was detected by the PreTect HPV-Proofer assay (NorChip AS, Klokkarstua, Norway). The presence of HPV DNA was investigated by Gp5+/6+ consensus PCR, and HPV DNA positive samples were then subjected to type specific PCR for the same 5 HPV types.

Results:

A total of 32.6% (n=92) samples were positive for HPV DNA by Gp5+/6+ PCR, and 24.8% (n=70) were found to be of types 16, 18, 31, 33, and 45. E6/E7 mRNA from the same five HPV types was observed in only 15.2% (n=43) of the cases.
The five carcinogenic HPV types 16, 18, 31, 33, and 45 accounted for 76% (70/92) of the HPV DNA positive samples, while an E6/E7 mRNA expression was detected in 61% (43/70) of these cases.
A cytological positive result was obtained in 8/282 cases (2.8%), of which ASCUS was observed in 5/8 cases and HPV condyloma in 3/8 cases. For the ASCUS cases, HPV DNA was detected by Gp5+/6+ consensus PCR and type specific PCR in 4/5 cases, whereas only one sample was found to contain HPV mRNA. For the HPV condyloma cases, however, HPV DNA was detected by Gp5+/6+ consensus PCR in all the samples (n=3), and by type specific PCR in 2/3 cases, while HPV E6/E7 mRNA expression was detected by PreTect HPV-Proofer in only 1 case.

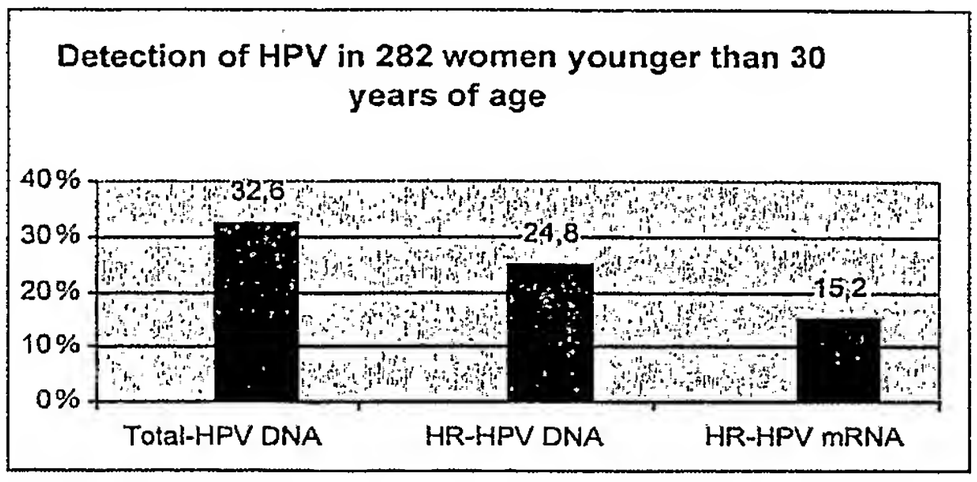
Discussion:

The presence of HPV in women younger than 30 years of age is higher than for older women (Molden et al, in preparation). This is also the case for the prevalence of the five carcinogenic HPV types 16, 18, 31, 33, and 45 compared to other types. Lack of E6/E7 transcripts may reflect an episomal state of the virus and hence a controlled regulation of the transcription process. These infections may be more likely to clear out. Integration of the virus, however, may disrupt the E2 gene, and thereby also its function as regulator of E6/E7 transcription.

Conclusion:

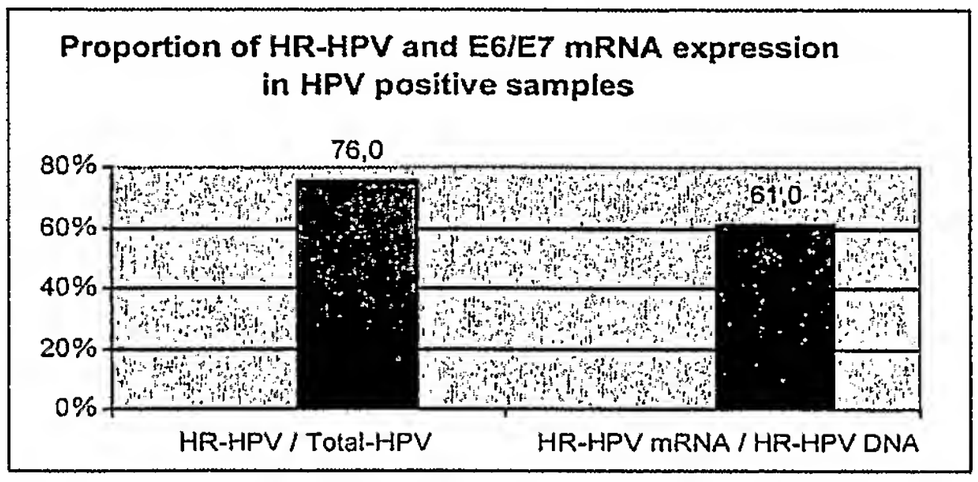
In this young outpatient population, HPV infection with an oncogene expression, is detected in less than 50% of the consensus PCR positive samples. Thus, monitoring E6/E7 gene expression for HPV types 16, 18, 31, 33 and 45 may be a valuable diagnostic test in addition to cytology. The relevance of either detection method will be studied by clinical follow-up of the women. Our opinion is that mRNA detection will be more discriminatory for progressive disease in young women.

Fig 1: Detection of HPV in women younger than 30 years of age



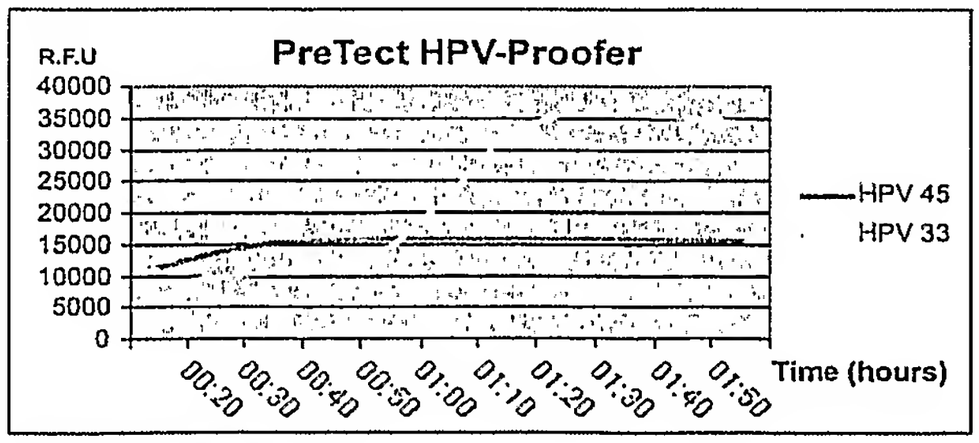
HR-HPV = High Risk HPV and includes HPV types 16, 18, 31, 33, and 45. DNA was detected by consensus and type specific PCR and mRNA was detected by the PreTect HPV-Proofer assay that utilises real-time multiplex NASBA.

Fig 2: Proportion of HR-HPV and E6/E7 mRNA expression in HPV positive samples



HR-HPV = High Risk HPV and includes HPV types 16, 18, 31, 33, and 45. DNA was detected by consensus and type specific PCR and mRNA was detected by the PreTect HPV-Proofer assay that utilises real-time multiplex NASBA.

Fig 3: Amplification of HPV 33 mRNA by the PreTect HPV-Proofer assay in sample 2755 diagnosed as HPV/condyloma.



R.F.U = Relative Fluorescence Units

Picture 1: Cytological HPV/condyloma

E6 and E7 mRNA expression from carcinogenic Human papillomavirus (HPV) in 4136 cervical samples collected from an outpatient population

Molden T[1], Kraus I[1] , Karlsen F[2], Skomedal H[2], Nygård J. F[3], Hagmar B[1]
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Objective:

High-risk human papillomavirus (HPV) infection is the main cause of cervical carcinoma and HPV DNA has been detected in more than 90% of cervical carcinoma samples, with the most common HPV types identified as HPV 16, 18, 31, 33, and 45. Naturally, it is tempting to include HPV detection and typing in screening programs in order to increase sensitivity. However, due to the high prevalence of HPV infections in normal samples the number of HPV DNA positive results may be to high for screening purposes.

Assays currently in use for HPV detection are based on L1 or E1 DNA amplification or hybridisation. However, the actual oncogenic effect is dependent on continuous expression from the E6 and E7 genes for successful transformation and maintenance of a neoplastic phenotype of the cervical carcinoma cells. Therefore, monitoring HPV mRNA expression in cervical smears may provide an accurate and informative diagnostic approach in cervical screening in addition to cytology.

The aim of this study was to identify the presence of E6/E7 mRNA and DNA in cytological HGSIL/CIN3 samples confirmed by histology.

Material and Methods:

The samples were collected from a well-screened outpatient population, including women older than 30 years of age (n=4136). E6/E7 transcripts from each of the high-risk HPV types 16, 18, 31, 33, and 45 were detected by the PreTect HPV-Proofer assay (NorChip AS, Klokkarstua, Norway), based on real-time multiplex NASBA. The presence of HPV DNA was investigated by Gp5+/6+ consensus PCR, and HPV DNA positive samples were then subjected to type specific PCR for HPV types 16, 18, 31, 33 and 45. Women with a cytological HGSIL diagnosis were referred to biopsy and histology. Histologically confirmed cases were registered at the Norwegian Cancer Registry. In Norway, cytological HGSIL can be divided into HGSIL/AGUS, HGSIL/ASC-H, HGSIL/CIN2, and HGSIL/CIN3.

Results:

Table 1:
Presence of HPV and E6/E7 mRNA in cytological HGSIL and histological CIN2+ samples

HPV positive by	Cytological HGSIL (n=25)	Histological CIN2+ (n=14)	Cytological HGSIL not verified by Histology (n=11)
PreTect HPV-Proofer	13 (52%)	12 (86%)	1 (9%)
Gp5+/6+ PCR	16 (64%)	13 (93%)	3 (27%)

In Norway, cytological HGSIL can be devided into HGSIL/AGUS, HGSIL/ASC-H, HGSIL/CIN2 and HGSIL/CIN3.

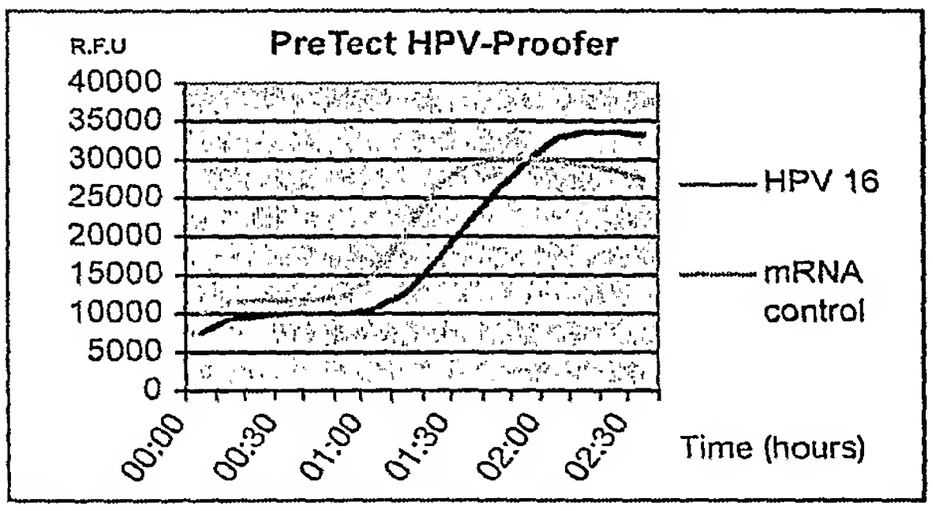
Results from HPV testing are summarized in **Table 1**. Of 25 cytological HGSIL cases, 14 were by histology confirmed as CIN2+. Two histological CIN2+ cases were by cytology diagnosed as HGSIL/ASC-H and HGSIL/CIN2. PreTect HPV-Proofer detected 52% (13/25) of the cytological HGSIL cases, 86% (12/14) of the histological CIN2+ cases, and 9% (1/11) of the cytological HGSIL cases not verified by histology. The numbers for Gp5+/6+ PCR are 64% (16/25), 93% (13/14), and 27% (3/11), respectively. The one histological CIN2+ sample positive by consensus PCR, yet negative by PreTect HPV-Proofer, was identified as HPV 35. The prevalence of HGSIL/CIN3 was 0.29% (12/4136) and the prevalence of histological CIN2+ was 56% (14/25).
Sensitivity, specificity, and positive and negative predictive values for PreTect HPV-Proofer and consensus PCR, are given in **Table 2**.

Table 2: *Sensitivity, specificity,* PPV and NPV for Pretect HPV-Proofer and Consensus PCR

	Total (n=4136) Endpoint cytological HGSIL/CIN3		Cytological HGSIL (n=25)* Endpoint histological CIN2+	
	PreTect HPV-Proofer	Consensus PCR	PreTect HPV-Proofer	Consensus PCR
Sensitivity	75,0%	83,3%	85,7%	92,9%
Specificity	97,2%	89,9%	90,9%	72,7%
PPV	7,3%	2,3%	92,3%	81,3%
NPV	97,2%	99,9%	83,3%	88,9%

* *Histology has only been performed on cytological HGSIL cases.* PPV=Positive Predictive Value. NPV=Negative Predicive Value.

Fig 1: E6/E7 mRNA amplification by real-time multiplex NASBA

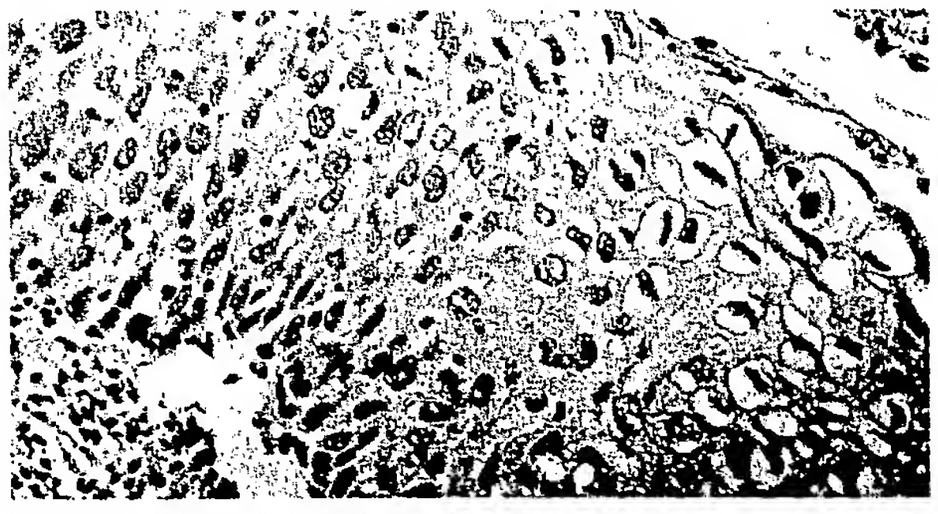


R.F.U = Relative Fluorescence Units

Discussion and Conclusion:

There was a good agreement between cytological HGSIL/CIN3 and histological CIN2+ cases, with only one cytological HGSIL/CIN3 not confirmed as CIN2+ by histology. In histologically verified CIN2+ cases, the detection grade for both HPV DNA and mRNA were high, and nearly identical. In cytological HGSIL cases not verified by histology, the detection grade for PreTect HPV-Proofer was lower than for consensus PCR. Together, cytological HGSIL/CIN3 and PreTect HPV-Proofer detected all histological CIN2+.
In conclusion, HPV E6/E7 transcripts from the five most frequently found carcinogenic HPV types, HPV 16, 18, 31, 33, and 45, seem to be present in nearly all histological CIN2+ cases.
The high specificity and positive predictive value for PreTect HPV-Proofer may be an advantage in HPV diagnostic and hence mRNA detection is a suitable supplement to cytology and histology.

Fig 2: Histological section of a CIN3 lesion.



HPV related cell changes (koilocytosis) are evident in the upper layers.

The potential of RNA as a target for molecular diagnostics in cervical carcinoma screening. A Review.

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Microarray technology and varied amplification methods have shown that RNA is valid as a target for routine molecular diagnostics and for future point-of-care testing. Indeed mRNA expression profiling which can simultaneously monitor 100 000 genes, both cellular and viral presents us with a new area of possibilities for the design of analytical tools to identify the disease state and disease susceptibility. National primary screening has resulted in a dramatic decrease in the incidence of cervical carcinoma. However, cytology is a subjective microscopic interpretation with poor reproducibility with a consequent trade-off between diagnostic sensitivity and specificity. The variable performance of cytology leads to marked variation in screening and management practices (Jenkins et al., 2003). Therefore, there is a great need for a more objective and cost-effective screening method with improved accuracy. It has been suggested that cytological smears may be more representative for molecular diagnostics of cell abnormalities than histocolposcopy. Detection of human papillomavirus targets may lend more accuracy and objectivity to the diagnostic process yet consensus DNA-HPV detection methods have been criticised due to their lack of clinical specificity and positive predictive value.

Table 1: Identification of E6/E7 mRNA from oncogenic human papillomavirus (HPV) in 41 cervical samples using the Pretect HPV-Proofer kit (1).

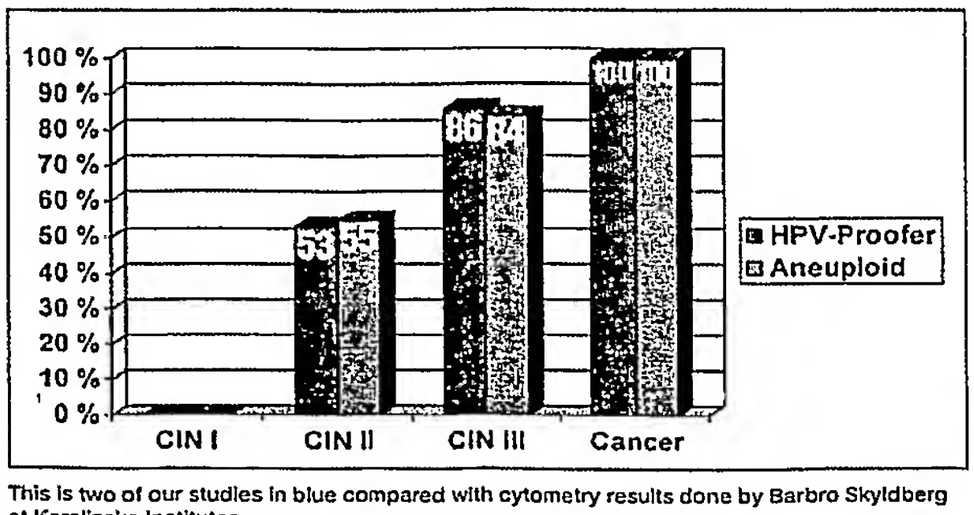
	Normal	ASCUS	Condy -loma	CIN1	CIN2	CIN3	SCC	Total
HPV mRNA	95 2%	12 21%	6 32%	0 0%	2 40%	9 75%	1 100%	126 3%
HPV DNA	361 9%	27 47%	14 74%	1 100%	2 40%	10 83%	1 100%	429 10%
Cytology	3970	57	19	1	5	12	1	4136*

*Results from groups unsatisfactory (64), AGUS (5) and ASC -H (2) not

Introduction

DNA is the part of the inheritated material that is passive target or coding sequence for RNA and Protein molecules that are going to perform the work in a biological system.
The main challenge using protein as a target for routine diagnostics has been low sensitivity, reproducibility and specificity. The main challenge using DNA as target for routine diagnostics has been lack of information about biological or clinical activity. However, RNA as a target for routine diagnostics may give the information of clinical activity, regulation or processes in addition to higher or equal sensitivity, reproducibility and specificity to DNA.
Organized screening programs based on periodic conventional Papaniocolaou (Pap) smears have been successful in reducing cervical cancer deaths in some few countries. In recent years, however, cervical cytological screening has come under attack because of a growing awareness of the test's imperfections, including irreproducibility and false negativity. Cervical cytological screening is entirely dependent on the combined judgment

Fig 2:
Rate of oncogene expression and DNA ploidy aberrations in biopsies (Skyldberg et al., 1999).



This is two of our studies in blue compared with cytometry results done by Barbro Skyldberg at Karolinska Institutes

Transcription from the HPV genome during the cervical carcinogenesis

Many studies claim that E6 and E7 splices or unspliced transcripts are produced on a very low level. None of these studies have performed studies of a large number of cytological normal cases. Three studies associated with us has shown that no transcripts are detected in around 50% of cytological normal samples indicate that E6 and E7 mRNA are not produced from samples with carcinogenic HPV DNA. It is tempting to define carcinogenic HPV DNA without oncogene expression as a part of the normal microbiological flora and not as an infection. When the HPV DNA has managed to enter the lower epithelial cell layer and has lost the regulation of E6 and E7 transcription and translation it may be defined as a real infection. The loss of regulation is not a part of the normal life cycle of the virus and it is therefore irreversible. In a two years follow-up study in Great Britain 21% of the samples with both HPV DNA and RNA at baseline loss their E6 and E7 transcript but not their DNA after two years (Kate et al., 2003, in press). The important question is weather these samples would retain their DNA and again turn on the E6 and E7 transcripts or loose the regulation of E6 and

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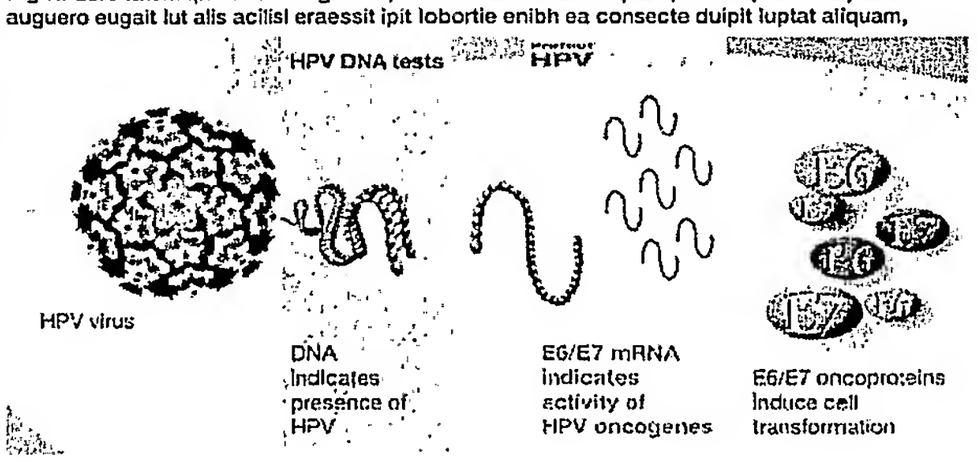
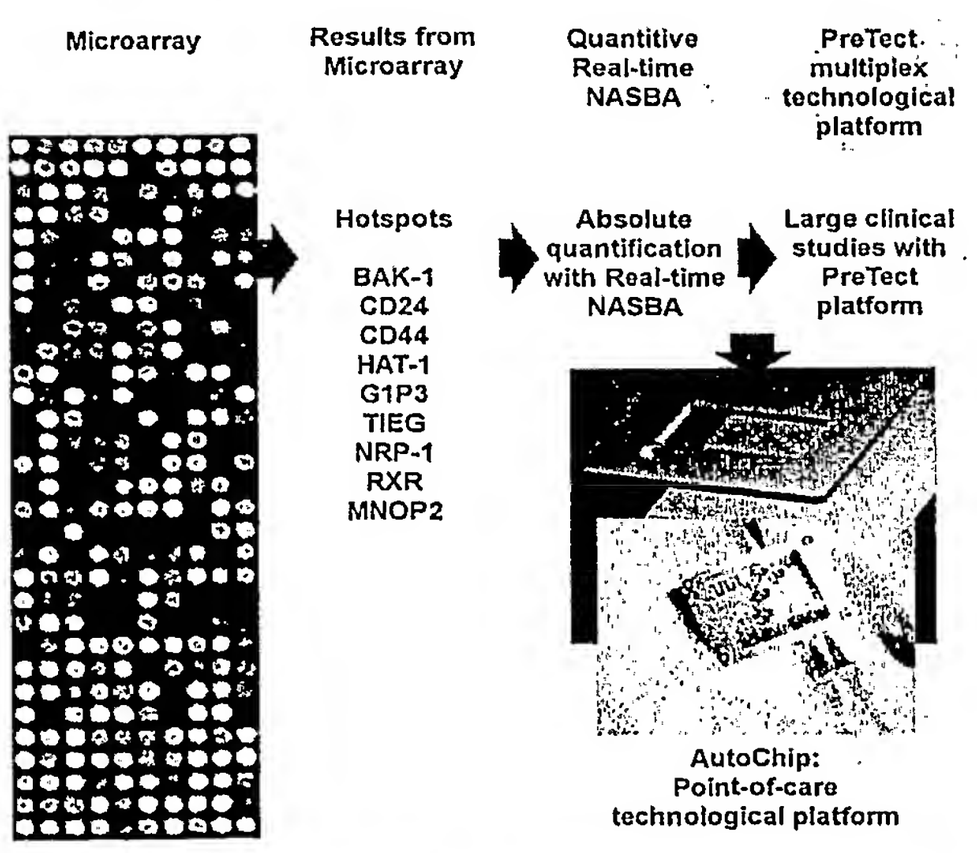


Fig 3: Lore tatem ipit autem auglamet,Lore tis numsandit aliquam, consequisim vulput la auguero eugait lut alis acilisl eraessit ipit lobortie enibh ea consecte duipit luptat aliquam,



References
Jenkins et al., Opt. Int. Dis 2001, 14:53-62; Tarkowski et al., Mole Diagn 2001, 6:125-30; Melchert et al., sub. 2003; Kraus et al., sub. 2003; Jiang and Milner Oncogene 2002, 21:6041-48; von Knebel et al., Eur J Cancer 2002, 38: 2229-42; Stacey et al., J. Virol 1995, 69:7023-30; Munoz et al., NEJM 2003, 348:518-27; Clifford et al., BJC 2003, 88: 63-73; Cuschieri et al., sub 2003.

Human Papillomavirus Type Specific DNA and RNA Persistence - Implications for Cervical Disease Progression and Monitoring

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Specialist Virology Centre, Royal Infirmary of Edinburgh, Edinburgh, Scotland

Abstract

In 2000, we monitored the course and persistence of human papillomavirus (HPV) infection in54 women who were HPV positive and free of any cytological disease using HPV-DNA genotyping with a linear array assay (baseline).

The impact of HPV infection on development of cervical cytological abnormality (dyskaryosis) was monitored by repeat HPV genotyping and cytological assessment 2 years later. Detection of mRNA transcripts of known HPV oncogenes E6 and E7 using NASBA methodology and specific molecular beacons for five common HPV types was also performed at both time points. A total of 11/54 (20%) women developed dyskaryosis after 2 years with 31/54 and 23/54 women exhibiting transient and persistent infections respectively, as monitored by DNA genotyping. Women who maintained type-specific persistent HPVinfection were significantly more likely to develop dyskaryosis compared to those who exhibited a transient infection ($P<0.001$). The presence of HPV mRNA E6/E7 transcripts was less sensitive but more specific for the detection of disease at follow up. Moreover, women who were DNA positive and also positive form RNA transcripts at baseline were significantly more likely to harbour persistent infection compared to those in whom DNA only was detected at baseline ($P<0.013$). This study highlights the importance of detecting persistent type specific HPV infection to identify those women more at risk of developing cervical abnormalities, either by repeated DNA genotyping, or potentially by RNA based techniques that may be more predictive of persistent infection if performed at a single time point.

TABLE II. Proportion of Detectable Persistent HPV Infections in Individuals With and Without Concurrent Evidence of Dyskaryosis on Follow-Up as Detected by DNA Genotyping and HPV RNA Transcript Detection

Cytological assessment on follow-up	No. of cases	No. of persistent infections (DNA)	No. of persistent infections (RNA)	No. of persistent infections (DNA or RNA)
Abnormal	11	10 (90.1)	6 (54.5)	10 (90.1)
Normal	43	13 (30.2)	5 (11.6)	15 (34.8)

TABLE III. Comparison of DNA Genotyping and RNA Transcript Detection for the Detection of 11 Cases of Dyskaryosis

Method of detection	Sensitivity		Specificity	
DNA	10/11	90.9%	19/43	44.2%
RNA	8/11	72.7%	35/43	81.4%

Serial biopsies from women with CIN and hr HPV oncogene expression in progressive lesions

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Background and design of the study

All women with birth/spontaneous abortion in 1996-7 were identified from the Medical Birth Registry of Norway. Based on the date of birth and last menstruation, (if reported) pregnancy related periods were specified for 116,547 women. The cohort was linked with the Cancer Registry data file, to identify women with abnormal Pap-smear during pregnancy. Women with (cervical intraepithelial lesion grade II + (CIN II+) cytological diagnoses (n=485), and women with biopsy during antenatal period (n=315) were identified. Altogether 253 women had follow-up biopsies taken after mean time of 301 days. Based on the change of the CIN grading in consecutive cervical histological diagnoses registered in the Norwegian Cancer Registry files, lesions were classified as progressive, regressive or persistent (Table 1).

nature	N	time of follow-up day	
		Total time	Mean time
Progression	44	24,224	550
Regression	29	12,063	416
Persistent	70	17,390	248
1sample	26		

Table 1. Distribution of the women by the nature of the lesion, defined as change in CIN grading in consecutive cervical histological diagnoses, registered in the Norwegian Cancer Registry files

Results

HPV analyses and pathology review of the "learning set" of 29 women with 69 consecutive visits are available. Blocks included into the learning set varied in respect to time of storage from min. 3.3 years to max. 8.2 years. The effect of storage time on HPV mRNA detection was assessed. Archival material, tested positively for HPV mRNA were stored for an overall shorter period of time (half a year). However, this difference was not significant statistically (Levene's test p=0.107).

Overall agreement between original and review pathology diagnoses (N=69) was low, Kappa=0.48. One of 5 normal biopsies, one of 10 CIN I and one of 15 CIN II lesions were tested positive for HPV 16 mRNA. Out of 38 CIN III lesions 21 (55.3%) were tested positive for HPV 16 mRNA. None tested positive for HPV 18, 31, 33 or 45.

When classifying lesion by their nature (Table 2), we took into account histology diagnoses of the study pathologist, time between two biopsies and Pap-smear history of women (from the Cytology Registry database). Examples of the variation of the morphology diagnoses for the progressive lesion are given in Figure 1a, b, c. Risk for progression among HPV16 mRNA positives: OR=12 (95% CI 0.5-280)

Nature	N	HPV16 mRNA	Mean fu-time (days)
Progressive	5	4 (80%)	430
Not progressive			
Regressive	4	1 (25%)	234
Persistence >1year	2	2 (100%)	445
Persistence <1year	14	6 (43%)	205
Progressive/Regressive	4	-	820

Table 2. Distribution of the women in the learning set by the nature of the lesion and HPV 16 mRNA positivity

Objective of the study

To estimate the prognostic value of HR-HPV mRNA in CIN for subsequent progression or regression of the disease.

Subjects

Legitimization of law about Biobanks requires contact with each person in order to obtain informed consent (IC). Out of 309 women involved, 70% gave IC. The final number of received paraffin blocks with cervical scrapes/biopsies/cones/hysterectomics were 1,484, taken from 128 women.

Material and Method

Original histology slides were revised by the study pathologist and one block with lesion per visit was selected for further analyses. All pathological and HPV analyses were done blinded. Five µm thick paraffin section, adjacent to the H&E staining was used for CIN grade assessment and following ten 5 µm paraffin sections were used for HPV analyses. Last 5µm paraffin section, adjacent to the H&E staining was again used for CIN grade assessment to ensure that material for HPV analyses were taken from the lesion. Before each case block, an empty paraffin block was used to control any possible cross-contamination. After deparaffinisation and extraction of DNA and RNA, real-time NASBA was used to amplify and detect full-length E6/E7 mRNA from HPV16, 18, 31, 33 and 45. Several different human mRNA was used as targets for performance control. Artificial oligonucleotides were used as positive controls.

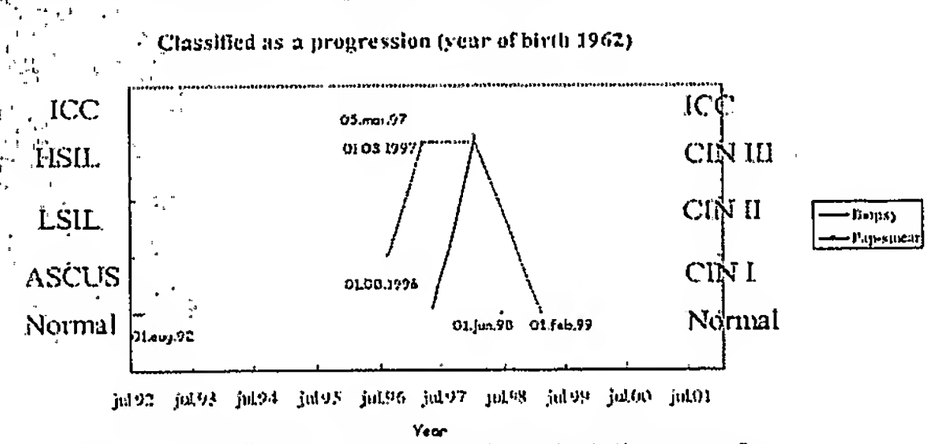

Figure 1a. Distribution of the morphological diagnoses for a case categorized as progressive

Figure 1b. Normal epithelium of a progressive lesion

Figure 1c. A CIN III section taken from a cervical cone with progressive lesion

Conclusions

CIN, while diagnosed during pregnancy opens the time-window for follow-up study of women who are not immediately treated. The Norwegian infrastructure made it possible to identify such women and to receive archive material for follow-up studies. Time of the storage did not have statistically significant effect on HPV 16 mRNA amplification and detection. The risk factors for CIN progression and morphological criteria for stratifying lesions into progressive and not progressive needs more clear definition. HPV16 mRNA was discovered more frequently in lesions with progression then in lesions with persisting morphology.

CLAIMS

1. An *in vitro* method of screening human subjects for the presence of human papillomavirus in at least one cell or tissue, wherein the human papillomavirus exhibits loss of regulation of E6/E7 mRNA expression and loss of replication, the method comprising detecting the presence of mRNA transcripts of the E6/E7 gene of a human papillomavirus in a test sample comprising mRNA derived from the cell or tissue, wherein the presence of E6/E7 mRNA transcripts in the sample is taken as an indication of the presence of human papilloma virus exhibiting loss of regulation of E6/E7 mRNA expression and loss of replication in the cell or tissue.

2. An *in vitro* method of screening human subjects for the presence of cellular changes characterized by enlarged cell nuclei and cellular aneuploidy in at least one cell or tissue, which method comprises detecting the presence of mRNA transcripts of the E6/E7 gene of human papillomavirus in a test sample comprising mRNA derived from the cell or tissue, wherein the presence of E6/E7 mRNA transcripts in the sample is taken as an indication that the cell or tissue under test exhibits the cellular changes.

3. A method according to claim 1 or claim 2 which comprises detecting expression of full length mRNA transcripts of the E6/E7 gene.

4. A method according to claims 1 to 3 which comprises detecting the presence of mRNA transcripts of the E6/E7 gene of human papillomavirus using a technique which is able to detect E6/E7 mRNA from at least one cancer-associated HPV type.

5. A method according to claim 4 which

comprises detecting the presence of mRNA transcripts of the E6/E7 gene of human papillomavirus using a technique which is able to detect E6/E7 mRNA from HPV types 16, 18, 31, 33, and preferably 45.

6. A method according to claim 4 which comprises detecting expression of mRNA transcripts of the E6/E7 gene from any one or more of HPV types 16, 18, 31, 33 or 45, wherein the presence of mRNA transcripts of the E6/E7 gene of human papillomavirus from any one of the tested HPV types in the sample is taken as a positive result.

7. A method according to any one of claims 1 to 6 wherein detection of expression of mRNA transcripts of the E6/E7 gene is carried out using an amplification reaction to amplify of a region of the mRNA, together with real-time detection of the products of the amplification reaction.

8. A method according to claim 7 wherein detection of expression of mRNA transcripts of the E6/E7 gene is carried out using real-time NASBA.

9. A method according to claim 8 wherein detection of expression of mRNA transcripts of the E6/E7 gene is carried out using the Pre-Tect HPV-Proofer™ assay kit.

10. A method according to any one of claims 1 to 9 wherein the human subjects are subjects previously identified as infected with human papillomavirus DNA, preferably in the cell or tissue under test.

11. A method according to any one of claims 1 to 10 wherein the human subjects are subjects having a previous diagnosis of ASCUS, CIN 1 lesions or condyloma.

12. An *in vitro* method of screening human

subjects for the presence of persistent infection with human papillomavirus, which method comprises screening the subject for expression of mRNA transcripts of the E6/E7 gene of human papillomavirus, wherein subjects positive for expression of mRNA transcripts of the E6/E7 gene of human papillomavirus are scored as having a persistent infection with human papillomavirus.